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DRUG TARGET ISOGENES:

POLYMORPHISMS IN THE INTERLEUKIN 4 RECEPTOR ALPHA GENE

*RELATED APPLICATIONS

This application is a division of PCT/US00/19094, filed July 13,2000.

FIELD OF THE INVENTION

This invention relates to variation in genes that encode pharmaceutically important proteins. In particular, this invention provides genetic variants of the human Interleukin 4 Receptor Alpha (IL4R α) gene and methods for identifying which variant(s) of this gene is/are possessed by an individual.

BACKGROUND OF THE INVENTION

Current methods for identifying pharmaceuticals to treat disease often start by identifying, cloning, and expressing an important target protein related to the disease. A determination of whether an agonist or antagonist is needed to produce an effect that may benefit a patient with the disease is then made. Then, vast numbers of compounds are screened against the target protein to find new potential drugs. The desired outcome of this process is a drug that is specific for the target, thereby reducing the incidence of the undesired side effects usually caused by a compound's activity at non-intended targets.

What this approach fails to consider, however, is that natural variability exists in any and every population with respect to a particular protein. A target protein currently used to screen drugs typically is expressed by a gene cloned from an individual who was arbitrarily selected. However, the nucleotide sequence of a particular gene may vary tremendously among individuals. Subtle alteration(s) in the primary nucleotide sequence of a gene encoding a target protein may be manifested as significant variation in expression of or in the structure and/or function of the protein. Such alterations may explain the relatively high degree of uncertainty inherent in treatment of individuals with drugs whose design is based upon a single representative example of the target. For example, it is well-established that some classes of drugs frequently have lower efficacy in some individuals than others, which means such individuals and their physicians must weigh the possible benefit of a larger dosage against a greater risk of side effects. In addition, variable information on the biological function or effects of a particular protein may be due to different scientists unknowingly studying different isoforms of the gene encoding the protein. Thus, information on the type and frequency of genomic variation that exists for pharmaceutically important proteins would be useful.

The organization of single nucleotide variations (polymorphisms) in the primary sequence of a gene into one of the limited number of combinations that exist as units of inheritance is termed a haplotype. Each haplotype therefore contains significantly more information than individual unorganized polymorphisms. Haplotypes provide an accurate measurement of the genomic variation in the two chromosomes of an individual.

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It is well-established that many diseases are associated with specific variations in gene sequences. However while there are examples in which individual polymorphisms act as genetic markers for a particular phenotype, in other cases an individual polymorphism may be found in a variety of genomic backgrounds and therefore shows no definitive coupling between the polymorphism and the causative site for the phenotype (Clark AG et al. 1998 Am J Hum Genet 63:595-612; Ulbrecht M et al. 2000 Am J Respir Crit Care Med 161: 469-74). In addition, the marker may be predictive in some populations, but not in other populations (Clark AG et al. 1998 supra). In these instances, a haplotype will provide a superior genetic marker for the phenotype (Clark AG et al. 1998 supra; Ulbrecht M et al. 2000, supra; Ruaño G & Stephens JC Gen Eng News 19 (21), December 1999).

Analysis of the association between each observed haplotype and a particular phenotype permits ranking of each haplotype by its statistical power of prediction for the phenotype. Haplotypes found to be strongly associated with the phenotype can then have that positive association confirmed by alternative methods to minimize false positives. For a gene suspected to be associated with a particular phenotype, if no observed haplotypes for that gene show association with the phenotype of interest, then it may be inferred that variation in the gene has little, if any, involvement with that phenotype (Ruaño & Stephens 1999, *supra*). Thus, information on the observed haplotypes and their frequency of occurrence in various population groups will be useful in a variety of research and clinical applications.

One possible drug target for the treatment of allergies, asthma, and other immune responses is the Interleukin 4 Receptor Alpha (IL4Rα) gene or its encoded product. IL4R is a transmembrane complex composed of two different protein subunits, a 140-kDa high affinity binding subunit named interleukin-4 receptor α (IL-4Rα; also known as CD124 antigen) and either a gamma-c subunit, which is present in several cytokine receptors, or an interleukin-13 receptor 1 (IL-13R1) subunit. Both subunits of the IL-4R are required to bind interleukin-4 (IL-4) and to mediate its transcription-activating effects through the tyrosine kinases, Jak1 and Jak3. Upon binding of IL-4 to the IL-4R, Jak 1 and Jak 3 phosphorylate the IL-4Rα subunit, creating binding sites in the cytoplasmic domain for many other proteins, including SOS, Stat-6, c-fes, and *src* homology phosphatase 1 (SHP-1). Activated Jak proteins also phosphorylate Stat-proteins, which travel into the nucleus and function as transcription factors. Other IL-4 signal pathways exist, but are less well characterized.

IL-4, one of the most important cytokines involved in the allergic response, is produced when cells from the immune system, in particular T cells, are activated in response to an allergen. Regulation of the immune response involves Helper T-cells that differentiate into two subtypes, Th1 and Th2. Th1 cells express interferon-gamma and interluekin-2 (IL-2), and mediate a cell-based immunity, where macrophages and neutrophils are prominently involved. Th1 cells also direct the IgE-producing B cells, as well as mast cells, basophils and eosinophils. Th2 cells produce IL-4, IL-5, IL-6, IL-10 and IL-13. Each Th cell subtype represses the other, so the immune system is forced into differentiation into either a Th1 or Th2 response against an external allergic challenge. In many instances an aberration of this response can render a pathological state such as a Th2 response against ragweed.

IL-4 induces in B cells the synthesis of IgE type antibodies that recognize specific allergens. IgE binds to receptors on mast cells and basophils and mediates the early humoral (sub-chronic) response on the B-side of the immune system. If an allergen binds mast cell-attached IgE, the mast cell releases mediators like histamine, and the eicosanoid leukotrienes and prostaglandins products, some of which cause the familiar symptoms of an acute allergic reaction: swelling, itching, mucous, and reddening of the skin. Later in this process eosinophils and other inflammatory cells migrate to the site of inflammation. This later phase is important in asthma, because the eosinophils may instigate a more chronic inflammation which can adversely scar lung tissue. IL-4 is at least partly responsible for recruiting eosinophils, because it induces synthesis of specific adhesion molecules on the capillary endothelium, and stimulates expression of IL-5 and eotaxin. IL-5 leads to the development of a large number of eosinophils from precursor cells in the bone marrow, and eotaxin stimulates their migration into the lung tissue.

It has been proposed that inhibition of IL-4 activity would disarm the Th2 component of the immune system. This would then allow the immune system to develop a natural tolerance towards common allergens without the full acute response to the challenge. In this way, tolerance may be induced in many patients, similar to what is sometimes achieved with hyposensitization shots for allergy patients. Thus, substances that inhibit IL-4 production and/or its binding to the IL-4 receptor, may improve the therapy of allergies and asthma.

The gene for IL-4Rα has eleven exons encoding an 825 amino acid protein and spans over 24 kb of the short arm of chromosome 16 (16p12.1) (Pritchard et al., *Genomics* 10:801, 1991; GenBank Accession No. AC004525). A reference sequence for IL4R-α gene, which corresponds to the reverse complement of nucleotides 100020-71331 in the GenBank Accession No. AC004525, is shown in Fig. 1 (SEQ ID NO:1). Reference sequences for IL-4Rα mRNA (GenBank Accession No. NM_000418) and the encoded IL4Rα precursor protein (GenBank Accession No. P24394) are shown in Figs. 2 and 3, respectively (SEQ ID NOS:2 and 3). Significant features reported for the IL-4Rα precursor include: a signal peptide located between a.a. 1 and 25; an extracellular domain between a.a. 26 and 232; disulfide bonds between a.a. 34 and 44 and between a.a. 74 and 86; glycosylation sites at amino acids 53, 98, 128, 134, 176, and 209; a transmembrane region between a.a. 233 and 256; and a cytoplasmic domain between a.a. 257 and 825.

Recently, several studies have suggested that genetic polymorphisms in the IL4-R α gene are associated with genetic predisposition to atopy and/or elevated serum IgE. Mitsuyasu et al., reported that polymorphisms at codons 75 and 576 affect IL-4R function (*Nat. Genet.* 19:119-120, 1998). The IL-4R α allele with isoleucine at amino acid position 75 (Ile75) in the extracellular domain is more responsive to IL-4 than the allele with valine at that position (Val75) and is associated with atopic asthma but not with non-atopic asthma (Mitsuyasu et al., *supra*). Also, the allele with arginine at position 576 (Arg576) in the cytoplasmic domain exhibits higher receptor activity than the glutamine allele (Glu576) due to

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reduced binding by the Arg576 allele of a negative regulatory molecule, src homology phosphatase 1 (Imani et al., J. Biol Chem. 272:7927-7931, 1997). The Arg576 allele has a higher frequency in patients with allergic inflammatory disorders, including atopy (Khurana Hershey et al., New Eng. J. Med. 337:1720-1725, 1997). In a recent study, Hershey et al., (WO 00/34789) reported that the Arg576 allele is significantly associated with asthma. Studies showed that patients who were homozygous for this allele had about a 9-fold higher risk towards asthma and that two copies of Arg576 are associated in an increase in asthma prevalence and severity. Kanemitsu et al. recently reported that the presence of either the Ile75 or Arg576 variant is significantly associated with susceptibility for developing systemic lupus erythematosus (SLE), a Th2-dominant systemic autoimmune disorder (Arthritis Rheum. 42:1298-1300). Another variant IL-4R α allele that is reportedly associated with atopy susceptibility has proline rather than serine at position 503 (Kruse et al., Immunol. 96:365-371, 1999). Other IL-4Ra gene polymorphisms leading to amino acid changes in the cytoplasmic domain of the protein product have been identified at codons 400 (E400A), 431 (C431R) and 786 (S786P) (Kruse et al., supra; Deichmann et al., Biochem. Biophys. Res. Commun. 231:696-697, 1997). A polymorphism of guanine or adenine at a position corresponding to nucleotide 55328 in Figure 1 has also been reported as well as a polymorphism of cytosine or thymine at a position corresponding to nucleotide 55430 (Buetow et al.,

Because of the potential for polymorphisms in the IL4R α gene to affect the expression and function of the encoded protein, it would be useful to determine whether additional polymorphisms exist in the IL4R α gene, as well as how such polymorphisms are combined in different copies of the gene. Such information would be useful for studying the biological function of IL4R α as well as in identifying drugs targeting this protein for the treatment of disorders related to its abnormal expression or function.

SUMMARY OF THE INVENTION

1999, Nat Genet. 21:323-5).

Accordingly, the inventors herein have discovered 38 novel polymorphic sites in the IL4R α gene. These polymorphic sites (PS) correspond to the following nucleotide positions in the reverse complement of the indicated GenBank Accession Number: 32884 (PS1), 32903 (PS2), 32961 (PS3), 33135 (PS4), 35763 (PS6), 35770 (PS7), 35817 (PS8), 35905 (PS9), 35944 (PS10), 35958 (PS11), 37330 (PS12), 37473 (PS13), 37586 (PS14), 37591 (PS15), 37604 (PS16), 37644 (PS17), 37678 (PS18), 43446 (PS19), 43703 (PS20), 53008 (PS21), 53099 (PS22), 53153 (PS23), 53456 (PS25), 53507 (PS27), 53513 (PS28), 53915 (PS30), 53949 (PS32), 54237 (PS33), 54468 (PS34), 54611 (PS35), 54698 (PS36), 54700 (PS37), 54741 (PS38), 54780 (PS39), 55083 (PS40), 55142 (PS41), 55539 (PS44) and 55758 (PS45) in AC004525. The polymorphisms at these sites are adenine or guanine at PS1, cytosine or thymine at PS2, guanine or cytosine at PS4, cytosine or thymine at PS6, guanine or adenine at PS7, thymine or cytosine at PS8, cytosine or thymine at PS9, cytosine or thymine at PS10, guanine or adenine at PS11, guanine or adenine at PS12, cytosine or thymine at PS13, cytosine or thymine at PS14,

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guanine or adenine at PS15, adenine or thymine at PS16, cytosine or adenine at PS17, cytosine or thymine at PS18, guanine or adenine at PS19, thymine or cytosine at PS20, adenine or cytosine at PS21, cytosine or thymine at PS22, thymine or cytosine at PS23, guanine or thymine at PS25, cytosine or thymine at PS27, thymine or cytosine at PS28, cytosine or thymine at PS30, guanine or adenine at PS32, cytosine or thymine at PS33, thymine or guanine at PS34, thymine or cytosine at PS35, thymine or cytosine at PS36, thymine or cytosine at PS37, cytosine or thymine at PS38, cytosine or guanine at PS39, adenine or guanine at PS40, guanine or adenine at PS41, cytosine or thymine at PS44 and guanine or adenine at PS45. In addition, the inventors have determined the identity of the alternative nucleotides present at these sites, as well as at the previously identified sites at nucleotides 35749 (PS5), 53413 (PS24), 53505 (PS26), 53721 (PS29), 53941 (PS31), 55328 (PS42), and 55430 (PS43). It is believed that IL4Rα-encoding polynucleotides containing one or more of the novel polymorphic sites reported herein will be useful in studying the expression and biological function of IL4Rα, as well as in developing drugs targeting this protein. In addition, information on the combinations of polymorphisms in the IL4Rα gene may have diagnostic and forensic applications.

Thus, in one embodiment, the invention provides an isolated polynucleotide comprising a nucleotide sequence which is a polymorphic variant of a reference sequence for the IL4Ra gene or a fragment thereof. The reference sequence comprises SEQ ID NO:1 and the polymorphic variant comprises at least one polymorphism selected from the group consisting of guanine at PS1, thymine at PS2, thymine at PS3, cytosine at PS4, thymine at PS6, adenine at PS7, cytosine at PS8, thymine at PS9, thymine at PS10, adenine at PS11, adenine at PS12, thymine at PS13, thymine at PS14, adenine at PS15, thymine at PS16, adenine at PS17, thymine at PS18, adenine at PS19, cytosine at PS20, cytosine at PS21, thymine at PS22, cytosine at PS23, thymine at PS25, thymine at PS27, cytosine at PS28, thymine at PS30, adenine at PS32, thymine at PS33, guanine at PS34, cytosine at PS35, cytosine at PS36, cytosine at PS37, thymine at PS38, guanine at PS39, guanine at PS40, adenine at PS41, thymine at PS44, and adenine at PS45. In a preferred embodiment, the polymorphic variant comprises one or more additional polymorphisms selected from the group consisting of guanine at PS5, cytosine at PS24, cytosine at PS26, cytosine at PS29, guanine at PS31, adenine at PS42, and thymine at PS43. A particularly preferred polymorphic variant is a naturally-occurring isoform (also referred to herein as an "isogene") of the IL4R α gene. An IL4R α isogene of the invention comprises adenine or guanine at PS1, cytosine or thymine at PS2, guanine or thymine at PS3, guanine or cytosine at PS4, cytosine or thymine at PS6, guanine or adenine at PS7, thymine or cytosine at PS8, cytosine or thymine at PS9, cytosine or thymine at PS10, guanine or adenine at PS11, guanine or adenine at PS12, cytosine or thymine at PS13, cytosine or thymine at PS14, guanine or adenine at PS15, adenine or thymine at PS16, cytosine or adenine at PS17, cytosine or thymine at PS18, guanine or adenine at PS19, thymine or cytosine at PS20, adenine or cytosine at PS21, cytosine or thymine at PS22, thymine or cytosine at PS23, guanine or thymine at PS25, cytosine or thymine at PS27, thymine or cytosine at PS28, cytosine or thymine at PS30, guanine or adenine at PS32, cytosine or thymine at PS33, thymine or guanine at PS34, thymine or cytosine at PS35,

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thymine or cytosine at PS36, thymine or cytosine at PS37, cytosine or thymine at PS38, cytosine or guanine at PS39, adenine or guanine at PS40, guanine or adenine at PS41, cytosine or thymine at PS44 and guanine or adenine at PS45. The invention also provides a collection of IL4Rα isogenes, referred to herein as an IL4Rα genome anthology.

An IL4R α isogene may be defined by the combination and order of these polymorphisms in the isogene, which is referred to herein as an IL4R α haplotype. Thus, the invention also provides data on the number of different IL4R α haplotypes found in the above four population groups. This haplotype data is useful in methods for deriving an IL4R α haplotype from an individual's genotype for the IL4R α gene and for determining an association between an IL4R α haplotype and a particular trait.

In another embodiment, the invention provides a polynucleotide comprising a polymorphic variant of a reference sequence for an IL4R α cDNA or a fragment thereof. The reference sequence comprises SEQ ID NO:2 (Fig. 2) and the polymorphic cDNA comprises at least one polymorphism selected from the group consisting of thymine at a position corresponding to nucleotide 237, adenine at a position corresponding to nucleotide 244, cytosine at a position corresponding to nucleotide 291, thymine at a position corresponding to nucleotide 501, adenine at a position corresponding to nucleotide 554, cytosine at a position corresponding to nucleotide 939, thymine at a position corresponding to nucleotide 1242, thymine at a position corresponding to nucleotide 1293, cytosine at a position corresponding to nucleotide 1299, thymine at a position corresponding to nucleotide 1701, adenine at a position corresponding to nucleotide 1735, thymine at a position corresponding to nucleotide 2023, guanine at a position corresponding to nucleotide 2254 and cytosine at a position corresponding to nucleotide 2397. In a preferred embodiment, the polymorphic variant comprises one or more additional polymorphisms selected from the group consisting of guanine at a position corresponding to 223, cytosine at a position corresponding to nucleotide 1199, cytosine at a position corresponding to 1291, cytosine at a position corresponding to nucleotide 1507 and guanine at a position corresponding to 1737.

Polynucleotides complementary to these IL4R α genomic and cDNA variants are also provided by the invention.

In other embodiments, the invention provides a recombinant expression vector comprising one of the polymorphic genomic variants operably linked to expression regulatory elements as well as a recombinant host cell transformed or transfected with the expression vector. The recombinant vector and host cell may be used to express IL4R α for protein structure analysis and drug binding studies.

In yet another embodiment, the invention provides a polypeptide comprising a polymorphic variant of a reference amino acid sequence for the IL4Rα protein. The reference amino acid sequence comprises SEQ ID NO:3 (Fig. 3) and the polymorphic variant comprises at least one variant amino acid selected from the group consisting of threonine at a position corresponding to amino acid 82, histidine at a position corresponding to amino acid 185, isoleucine at a position corresponding to amino acid 579, serine at a position corresponding to amino acid 675, and alanine at a position corresponding to amino

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acid 752. In some embodiments, the polymorphic variant also comprises at least one variant amino acid selected from the group consisting of valine at a position corresponding to amino acid 75, alanine at a position corresponding to amino acid 400, arginine at a position corresponding to amino acid 431, proline at a position corresponding to amino acid 503, and arginine at a position corresponding to amino acid 576. A polymorphic variant of IL4R α is useful in studying the effect of the variation on the biological activity of IL4R α as well as studying the binding affinity of candidate drugs targeting IL4R α for the treatment of allergies, asthma, and other immune responses.

The present invention also provides antibodies that recognize and bind to the above polymorphic IL4R α protein variant. Such antibodies can be utilized in a variety of diagnostic and prognostic formats and therapeutic methods.

In other embodiments, the invention provides methods, compositions, and kits for haplotyping and/or genotyping the IL4R α gene in an individual. The methods involve identifying the nucleotide or nucleotide pair present at one or more polymorphic sites selected from PS1, PS2, PS3, PS4, PS6, PS7, PS8, PS9, PS10, PS11, PS12, PS13, PS14, PS15, PS16, PS17, PS18, PS19, PS20, PS21, PS22, PS23, PS25, PS27, PS28, PS30, PS32, PS33, PS34, PS35, PS36, PS37, PS38, PS39, PS40, PS41, PS44, and PS45 in one or both copies of the IL4R α gene from the individual. The compositions contain oligonucleotide probes and primers designed to specifically hybridize to one or more target regions containing, or that are adjacent to, a polymorphic site. The methods and compositions for establishing the genotype or haplotype of an individual at the novel polymorphic sites described herein are useful for studying the effect of the polymorphisms in the etiology of diseases affected by the expression and function of the IL4R α protein, studying the efficacy of drugs targeting IL4R α , predicting individual susceptibility to diseases affected by the expression and function of the IL4R α protein and predicting individual responsiveness to drugs targeting IL4R α .

In yet another embodiment, the invention provides a method for identifying an association between a genotype or haplotype and a trait. In preferred embodiments, the trait is susceptibility to a disease, severity of a disease, the staging of a disease or response to a drug. Such methods have applicability in developing diagnostic tests and therapeutic treatments for allergies, asthma, and other immune responses.

The present invention also provides transgenic animals comprising one of the IL4R α genomic polymorphic variants described herein and methods for producing such animals. The transgenic animals are useful for studying expression of the IL4R α isogenes *in vivo*, for *in vivo* screening and testing of drugs targeted against IL4R α protein, and for testing the efficacy of therapeutic agents and compounds for allergies, asthma, and other immune responses in a biological system.

The present invention also provides a computer system for storing and displaying polymorphism data determined for the IL4R α gene. The computer system comprises a computer processing unit; a display; and a database containing the polymorphism data. The polymorphism data includes the

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polymorphisms, the genotypes and the haplotypes identified for the IL4R α gene in a reference population. In a preferred embodiment, the computer system is capable of producing a display showing IL4R α haplotypes organized according to their evolutionary relationships.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 illustrates a reference sequence for the IL4Rα gene that is the reverse complement of part of Genbank Accession Number AC004525.1; contiguous lines; SEQ ID NO:1), with the underlines indicating the start and stop codons, shading indicating the reference coding sequence, and the bold nucleotides indicating the polymorphic sites and polymorphisms identified by Applicants in a reference population.

Figure 2 illustrates a reference sequence for the IL4Rα coding sequence (GenBank Accession Number X52425; contiguous lines; SEQ ID NO:2), with the underlines indicating the start and stop codons, and the bold nucleotides indicating the polymorphic sites and polymorphisms identified by Applicants in a reference population.

Figure 3 illustrates a reference sequence for the IL4Rα protein (GenBank Accession Number CAA36672; contiguous lines; SEQ ID NO:3), with the bold amino acids indicating the amino acid variations caused by the polymorphisms of Fig. 2.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention is based on the discovery of novel variants of the IL4Rα gene. As described in more detail below, the inventors herein discovered 38 novel polymorphic sites by characterizing the IL4Rα gene found in genomic DNAs isolated from Index Repository IA that contains immortalized cell lines from one chimpanzee and 93 human individuals and Index Repository IB that contains 70 human individuals. Theses two repositories contain 51 individuals in common.

The human individuals in Index Repository IA included a reference population of 79 unrelated individuals self-identified as belonging to one of four major population groups: Caucasian (22 individuals), African descent (20 individuals) Asian (20 individuals) Hispanic/Latino (17 individuals). To the extent possible, the members of this reference population were organized into population subgroups by the self-identified ethnogeographic origin of their four grandparents as shown in Table 1 below. In addition, Index Repository IA contains three unrelated indigenous American Indians (one from each of North, Central, and South America), one three-generation Caucasian family (From the CEPH Utah cohort) and one two-generation African-American family.

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Table 1. Population Groups in Index Repository IA

Population Group	Population Subgroup	No. of Individuals
African descent		20
	Sierra Leone	1
Asian		20
	Burma	1
	China	3
	Japan	6
	Korea	1
	Philippines	5
	Vietnam	4
Caucasian		22
	British Isles	3
	British Isles/Central	4
	British Isles/Eastern	1
	Central/Eastern	1
	Eastern	3
	Central/Mediterranean	1
	Mediterranean	2
	Scandinavian	2
Hispanic/Latino		17
	Caribbean	7
	Caribbean (Spanish Descent)	2
	Central American (Spanish Descent)	1
	Mexican American	4
	South American (Spanish Descent)	3

Index Repository IB contains a reference population of 70 human individuals comprised of 4 three-generation families (from the CEPH Utah cohort) as well as unrelated African-American, Asian, and Caucasian individuals. A total of 38 individuals in this reference population are unrelated.

Using the IL4R α genotypes identified in the Index Repositories and the methodology described in the Examples below, the inventors herein also determined the haplotypes found on each chromosome for most human members of this repository. The IL4 α genotypes and haplotypes found in the Index Repositories include those shown in Tables 4 and 5, respectively. The polymorphism and haplotype data disclosed herein are useful for studying population diversity, anthropological lineage, the significance of diversity and lineage at the phenotypic level, paternity testing, forensic applications, and for identifying associations between the IL4R α genetic variation and a trait such as level of drug response or susceptibility to disease.

In the context of this disclosure, the following terms shall be defined as follows unless otherwise indicated:

Allele - A particular form of a genetic locus, distinguished from other forms by its particular nucleotide sequence.

Candidate Gene - A gene which is hypothesized to be responsible for a disease, condition, or

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the response to a treatment, or to be correlated with one of these.

Gene - A segment of DNA that contains all the information for the regulated biosynthesis of an RNA product, including promoters, exons, introns, and other untranslated regions that control expression.

Genotype – An unphased 5' to 3' sequence of nucleotide pair(s) found at one or more polymorphic sites in a locus on a pair of homologous chromosomes in an individual. As used herein, genotype includes a full-genotype and/or a sub-genotype as described below.

Full-genotype – The unphased 5' to 3' sequence of nucleotide pairs found at all known polymorphic sites in a locus on a pair of homologous chromosomes in a single individual.

Sub-genotype – The unphased 5' to 3' sequence of nucleotides seen at a subset of the known polymorphic sites in a locus on a pair of homologous chromosomes in a single individual.

Genotyping - A process for determining a genotype of an individual.

Haplotype – A 5' to 3' sequence of nucleotides found at one or more polymorphic sites in a locus on a single chromosome from a single individual. As used herein, haplotype includes a full-haplotype and/or a sub-haplotype as described below.

Full-haplotype – The 5' to 3' sequence of nucleotides found at all known polymorphic sites in a locus on a single chromosome from a single individual.

Sub-haplotype – The 5' to 3' sequence of nucleotides seen at a subset of the known polymorphic sites in a locus on a single chromosome from a single individual.

Haplotype pair – The two haplotypes found for a locus in a single individual.

Haplotyping – A process for determining one or more haplotypes in an individual and includes use of family pedigrees, molecular techniques and/or statistical inference.

Haplotype data - Information concerning one or more of the following for a specific gene: a listing of the haplotype pairs in each individual in a population; a listing of the different haplotypes in a population; frequency of each haplotype in that or other populations, and any known associations between one or more haplotypes and a trait.

Isoform – A particular form of a gene, mRNA, cDNA or the protein encoded thereby, distinguished from other forms by its particular sequence and/or structure.

Isogene – One of the isoforms of a gene found in a population. An isogene contains all of the polymorphisms present in the particular isoform of the gene.

Isolated — As applied to a biological molecule such as RNA, DNA, oligonucleotide, or protein, isolated means the molecule is substantially free of other biological molecules such as nucleic acids, proteins, lipids, carbohydrates, or other material such as cellular debris and growth media. Generally, the term "isolated" is not intended to refer to a complete absence of such material or to absence of water, buffers, or salts, unless they are present in amounts that substantially interfere with the methods of the present invention.

Locus - A location on a chromosome or DNA molecule corresponding to a gene or a physical or

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phenotypic feature.

Naturally-occurring – A term used to designate that the object it is applied to, e.g., naturally-occurring polynucleotide or polypeptide, can be isolated from a source in nature and which has not been intentionally modified by man.

Nucleotide pair – The nucleotides found at a polymorphic site on the two copies of a chromosome from an individual.

Phased – As applied to a sequence of nucleotide pairs for two or more polymorphic sites in a locus, phased means the combination of nucleotides present at those polymorphic sites on a single copy of the locus is known.

Polymorphic site (PS) – A position within a locus at which at least two alternative sequences are found in a population, the most frequent of which has a frequency of no more than 99%.

Polymorphic variant – A gene, mRNA, cDNA, polypeptide or peptide whose nucleotide or amino acid sequence varies from a reference sequence due to the presence of a polymorphism in the gene.

Polymorphism – The sequence variation observed in an individual at a polymorphic site. Polymorphisms include nucleotide substitutions, insertions, deletions and microsatellites and may, but need not, result in detectable differences in gene expression or protein function.

Polymorphism data – Information concerning one or more of the following for a specific gene: location of polymorphic sites; sequence variation at those sites; frequency of polymorphisms in one or more populations; the different genotypes and/or haplotypes determined for the gene; frequency of one or more of these genotypes and/or haplotypes in one or more populations; any known association(s) between a trait and a genotype or a haplotype for the gene.

Polymorphism Database – A collection of polymorphism data arranged in a systematic or methodical way and capable of being individually accessed by electronic or other means.

Polynucleotide – A nucleic acid molecule comprised of single-stranded RNA or DNA or comprised of complementary, double-stranded DNA.

Population Group - A group of individuals sharing a common ethnogeographic origin.

Reference Population – A group of subjects or individuals who are predicted to be representative of the genetic variation found in the general population. Typically, the reference population represents the genetic variation in the population at a certainty level of at least 85%, preferably at least 90%, more preferably at least 95% and even more preferably at least 99%.

Single Nucleotide Polymorphism (SNP) – Typically, the specific pair of nucleotides observed at a single polymorphic site. In rare cases, three or four nucleotides may be found.

Subject – A human individual whose genotypes or haplotypes or response to treatment or disease state are to be determined.

Treatment - A stimulus administered internally or externally to a subject.

Unphased - As applied to a sequence of nucleotide pairs for two or more polymorphic sites in a

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locus, unphased means the combination of nucleotides present at those polymorphic sites on a single copy of the locus is not known.

The inventors herein have discovered 38 novel polymorphic sites , and confirmed the existence of 7 other sites, in the IL4Rα gene. The polymorphic sites identified by the inventors are referred to as PS1-45 to designate the order in which they are located in the gene (see Table 3 below), with the novel polymorphic site referred to as PS1, PS2, PS3, PS4, PS6, PS7, PS8, PS9, PS10, PS11, PS12, PS13, PS14, PS15, PS16, PS17, PS18, PS19, PS20, PS21, PS22, PS23, PS25, PS27, PS28, PS30, PS32, PS33, PS34, PS35, PS36, PS37, PS38, PS39, PS40, PS41, PS44, and PS45 and the previously reported polymorphic sites referred to as PS5, PS24, PS26, PS29, PS31, PS42, and PS43.

Thus, in one embodiment, the invention provides an isolated polynucleotide comprising a polymorphic variant of the IL4Rα gene or a fragment of the gene which contains at least one of the novel polymorphic sites described herein. The nucleotide sequence of a variant IL4Rα gene is identical to the reference genomic sequence for those portions of the gene examined, as described in the Examples below, except that it comprises a different nucleotide at one or more of the novel polymorphic sites PS1, PS2, PS3, PS4, PS6, PS7, PS8, PS9, PS10, PS11, PS12, PS13, PS14, PS15, PS16, PS17, PS18, PS19, PS20, PS21, PS22, PS23, PS25, PS27, PS28, PS30, PS32, PS33, PS34, PS35, PS36, PS37, PS38, PS39, PS40, PS41, PS44, and PS45, and may also comprise one or more additional polymorphisms selected from the group consisting of PS5, PS24, PS26, PS29, PS31, PS42, and PS43. Similarly, the nucleotide sequence of a variant fragment of the IL4Rα gene is identical to the corresponding portion of the reference sequence except for having a different nucleotide at one or more of the novel polymorphic sites described herein. Thus, the invention specifically does not include polynucleotides comprising a nucleotide sequence identical to the reference sequence (or other reported IL4Rα sequences) or to portions of the reference sequence (or other reported IL4Rα sequences), except for genotyping oligonucleotides as described below.

The location of a polymorphism in a variant gene or fragment is identified by aligning its sequence against SEQ ID NO:1. The polymorphism is selected from the group consisting of guanine at PS1, thymine at PS2, thymine at PS3, cytosine at PS4, thymine at PS6, adenine at PS7, cytosine at PS8, thymine at PS9, thymine at PS10, adenine at PS11, adenine at PS12, thymine at PS13, thymine at PS14, adenine at PS15, thymine at PS16, adenine at PS17, thymine at PS18, adenine at PS19, cytosine at PS20, cytosine at PS21, thymine at PS22, cytosine at PS23, thymine at PS25, thymine at PS27, cytosine at PS28, thymine at PS30, adenine at PS32, thymine at PS34, cytosine at PS35, cytosine at PS36, cytosine at PS37, thymine at PS38, guanine at PS39, guanine at PS40, adenine at PS41, thymine at PS44, and adenine at PS45. In a preferred embodiment, the polymorphic variant comprises a naturally-occurring isogene of the IL4Rα gene which is defined by any one of haplotypes 1-53 shown in Table 5 below.

Polymorphic variants of the invention may be prepared by isolating a clone containing the

IL4Rα gene from a human genomic library. The clone may be sequenced to determine the identity of the nucleotides at the polymorphic sites described herein. Any particular variant claimed herein could be prepared from this clone by performing *in vitro* mutagenesis using procedures well-known in the art.

IL4Rα isogenes may be isolated using any method that allows separation of the two "copies" of the IL4Rα gene present in an individual, which, as readily understood by the skilled artisan, may be the same allele or different alleles. Separation methods include targeted *in vivo* cloning (TIVC) in yeast as described in WO 98/01573, U.S. Patent No. 5,866,404, and copending U.S. application Serial No. 08/987,966. Another method, which is described in copending U.S. Application Serial No. 08/987,966, uses an allele specific oligonucleotide in combination with primer extension and exonuclease degradation to generate hemizygous DNA targets. Yet other methods are single molecule dilution (SMD) as described in Ruaño et al., Proc. Natl. Acad. Sci. 87:6296-6300, 1990; and allele specific PCR (Ruaño et al., 17 Nucleic Acids. Res. 8392, 1989; Ruaño et al., 19 Nucleic Acids Res. 6877-6882, 1991; Michalatos-Beloin et al., 24 Nucleic Acids Res. 4841-4843, 1996).

The invention also provides IL4R α genome anthologies, which are collections of IL4R α isogenes found in a given population. The population may be any group of at least two individuals, including but not limited to a reference population, a population group, a family population, a clinical population, and a same sex population. An IL4R α genome anthology may comprise individual IL4R α isogenes stored in separate containers such as microtest tubes, separate wells of a microtitre plate and the like. Alternatively, two or more groups of the IL4R α isogenes in the anthology may be stored in separate containers. Individual isogenes or groups of isogenes in a genome anthology may be stored in any convenient and stable form, including but not limited to in buffered solutions, as DNA precipitates, freeze-dried preparations and the like. A preferred IL4R α genome anthology of the invention comprises a set of isogenes defined by the haplotypes shown in Table 5 below.

An isolated polynucleotide containing a polymorphic variant nucleotide sequence of the invention may be operably linked to one or more expression regulatory elements in a recombinant expression vector capable of being propagated and expressing the encoded IL4R α protein in a prokaryotic or a eukaryotic host cell. Examples of expression regulatory elements which may be used include, but are not limited to, the lac system, operator and promoter regions of phage lambda, yeast promoters, and promoters derived from vaccinia virus, adenovirus, retroviruses, or SV40. Other regulatory elements include, but are not limited to, appropriate leader sequences, termination codons, polyadenylation signals, and other sequences required for the appropriate transcription and subsequent translation of the nucleic acid sequence in a given host cell. Of course, the correct combinations of expression regulatory elements will depend on the host system used. In addition, it is understood that the expression vector contains any additional elements necessary for its transfer to and subsequent replication in the host cell. Examples of such elements include, but are not limited to, origins of replication and selectable markers. Such expression vectors are commercially available or are readily

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constructed using methods known to those in the art (e.g., F. Ausubel et al., 1987, in "Current Protocols in Molecular Biology", John Wiley and Sons, New York, New York). Host cells which may be used to express the variant IL4Rα sequences of the invention include, but are not limited to, eukaryotic and mammalian cells, such as animal, plant, insect and yeast cells, and prokaryotic cells, such as E. coli, or algal cells as known in the art. The recombinant expression vector may be introduced into the host cell using any method known to those in the art including, but not limited to, microinjection, electroporation, particle bombardment, transduction, and transfection using DEAE-dextran, lipofection, or calcium phosphate (see e.g., Sambrook et al. (1989) in "Molecular Cloning. A Laboratory Manual", Cold Spring Harbor Press, Plainview, New York). In a preferred aspect, eukaryotic expression vectors that function in eukaryotic cells, and preferably mammalian cells, are used. Non-limiting examples of such vectors include vaccinia virus vectors, adenovirus vectors, herpes virus vectors, and baculovirus transfer vectors. Preferred eukaryotic cell lines include COS cells, CHO cells, HeLa cells, NIH/3T3 cells, and embryonic stem cells (Thomson, J. A. et al., 1998 Science 282:1145-1147). Particularly preferred host cells are mammalian cells.

As will be readily recognized by the skilled artisan, expression of polymorphic variants of the IL4R α gene will produce IL4R α mRNAs varying from each other at any polymorphic site retained in the spliced and processed mRNA molecules. These mRNAs can be used for the preparation of an IL4R α cDNA comprising a nucleotide sequence which is a polymorphic variant of the IL4R α reference coding sequence shown in Figure 2. Thus, the invention also provides IL4Ra mRNAs and corresponding cDNAs which comprise a nucleotide sequence that is identical to SEQ ID NO:2 (Fig. 2), or its corresponding RNA sequence, except for having one or more polymorphisms selected from the group consisting of thymine at a position corresponding to nucleotide 237, adenine at a position corresponding to nucleotide 244, cytosine at a position corresponding to nucleotide 291, thymine at a position corresponding to nucleotide 501, adenine at a position corresponding to nucleotide 554, cytosine at a position corresponding to nucleotide 939, thymine at a position corresponding to nucleotide 1242, thymine at a position corresponding to nucleotide 1293, cytosine at a position corresponding to nucleotide 1299, thymine at a position corresponding to nucleotide 1701, adenine at a position corresponding to nucleotide 1735, thymine at a position corresponding to nucleotide 2023, guanine at a position corresponding to nucleotide 2254 and cytosine at a position corresponding to nucleotide 2397, and may also comprise one or more additional polymorphisms selected from the group consisting of guanine at a position corresponding to 223, cytosine at a position corresponding to nucleotide 1199, cytosine at a position corresponding to 1291, cytosine at a position corresponding to nucleotide 1507 and guanine at a position corresponding to 1737. Fragments of these variant mRNAs and cDNAs are included in the scope of the invention, provided they contain the novel polymorphisms described herein. The invention specifically excludes polynucleotides identical to previously identified and characterized $IL4R\alpha$ cDNAs and fragments thereof. Polynucleotides comprising a variant RNA or DNA sequence

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may be isolated from a biological sample using well-known molecular biological procedures or may be chemically synthesized.

Genomic and cDNA fragments of the invention comprise at least one novel polymorphic site identified herein and have a length of at least 10 nucleotides and may range up to the full length of the gene. Preferably, a fragment according to the present invention is between 100 and 3000 nucleotides in length, and more preferably between 200 and 2000 nucleotides in length, and most preferably between 500 and 1000 nucleotides in length.

In describing the polymorphic sites identified herein, reference is made to the sense strand of the gene for convenience. However, as recognized by the skilled artisan, nucleic acid molecules containing the IL4R α gene may be complementary double stranded molecules and thus reference to a particular site on the sense strand refers as well to the corresponding site on the complementary antisense strand. Thus, reference may be made to the same polymorphic site on either strand and an oligonucleotide may be designed to hybridize specifically to either strand at a target region containing the polymorphic site. Thus, the invention also includes single-stranded polynucleotides which are complementary to the sense strand of the IL4R α genomic variants described herein.

Polynucleotides comprising a polymorphic gene variant or fragment may be useful for therapeutic purposes. For example, where a patient could benefit from expression, or increased expression, of a particular IL4R α protein isoform, an expression vector encoding the isoform may be administered to the patient. The patient may be one who lacks the IL4R α isogene encoding that isoform or may already have at least one copy of that isogene.

In other situations, it may be desirable to decrease or block expression of a particular IL4R α isogene. Expression of an IL4R α isogene may be turned off by transforming a targeted organ, tissue or cell population with an expression vector that expresses high levels of untranslatable mRNA for the isogene. Alternatively, oligonucleotides directed against the regulatory regions (e.g., promoter, introns, enhancers, 3' untranslated region) of the isogene may block transcription. Oligonucleotides targeting the transcription initiation site, e.g., between positions -10 and +10 from the start site are preferred. Similarly, inhibition of transcription can be achieved using oligonucleotides that base-pair with region(s) of the isogene DNA to form triplex DNA (see e.g., Gee et al. in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing Co., Mt. Kisco, N.Y., 1994). Antisense oligonucleotides may also be designed to block translation of IL4R α mRNA transcribed from a particular isogene. It is also contemplated that ribozymes may be designed that can catalyze the specific cleavage of IL4R α mRNA transcribed from a particular isogene.

The oligonucleotides may be delivered to a target cell or tissue by expression from a vector introduced into the cell or tissue *in vivo* or *ex vivo*. Alternatively, the oligonucleotides may be formulated as a pharmaceutical composition for administration to the patient. Oligoribonucleotides and/or oligodeoxynucleotides intended for use as antisense oligonucleotides may be modified to increase

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stability and half-life. Possible modifications include, but are not limited to phosphorothioate or 2' O-methyl linkages, and the inclusion of nontraditional bases such as inosine and queosine, as well as acetyl, methyl-, thio-, and similarly modified forms of adenine, cytosine, guanine, thymine, and uracil which are not as easily recognized by endogenous nucleases.

The invention also provides an isolated polypeptide comprising a polymorphic variant of the reference IL4R α amino acid sequence shown in 3. The location of a variant amino acid in an IL4R α polypeptide or fragment of the invention is identified by aligning its sequence against Fig. 3. An IL4R α protein variant of the invention comprises an amino acid sequence identical to SEQ ID NO: 3 except for having one or more variant amino acids selected from the group consisting of threonine at a position corresponding to amino acid 82, histidine at a position corresponding to amino acid 185, isoleucine at a position corresponding to amino acid 579, serine at a position corresponding to amino acid 675, and alanine at a position corresponding to amino acid 752, and may also comprise one or more additional variant amino acids selected from the group consisting of valine at a position corresponding to amino acid 75, alanine at a position corresponding to amino acid 400, arginine at a position correpsonding to amino acid 431, proline at a position corresponding to amino acid 503, and arginine at a position corresponding to amino acid 576. The invention specifically excludes amino acid sequences identical to those previously identified for IL4R α , including SEQ ID NO: 3, and previously described fragments thereof. IL4Ra protein variants included within the invention comprise all amino acid sequences based on SEQ ID NO: 3 and having the combination of amino acid variations described in Table 2 below. In preferred embodiments, an IL4Ra protein variant of the invention is encoded by an isogene defined by one of the observed haplotypes shown in Table 5.

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·		Table	2. Novel	Polymor	phic Var	iant of IL	4Rα			
Polymorphic										
Variant				Amino A	cid Posi	tion and	Identities			
Number	75	82	185	400	431	503	576	579	675	752
1.	75	A	R	E	C	S	Q	V	Р	Α
2		Â	R	E	C	S	à	V	S	S
			R	E	C	S	- ä -	V	S	Ā
3	<u> </u>	A			C	S	ä		P	S
4		A	R	E				1	P	A
5		Α	R	E	C)	S	Q			
6		Α	R	E	С	S	Q	<u> </u>	S	S
7	1	Α	R	E	C	S	Q		S	<u>A</u>
8	1	Α	R	E	C	S	R	V	Р	A
9	1	Α	R	E	С	S	R	V	S	S
10	ı	Α	R	E	С	S	R	V	S	Α
11	j	Α	R	Ε	С	S	R		Р	S
12	1	A	R	Ē	С	S	R		Р	Α
13	i	Ā	R	Е	С	S	R	·	S	S
14	i	A	R	E	C	S	R	<u> </u>	S	Α
	1	A	R	Ē	C	P	Q	V	P	A
15		A	R	E	C	Р	σα	V	S	S
16	1 1			E	C	P	Q	V	S	A
17	 	A	R	E	C	P	Q	V	P	s
18		Α	R							
19	1	Α	R	E	С	Р	Q	! :	Р	A
20	l l	Α	R	E	C	Р	Q		S	S
21	l	Α	R	E	С	Р	Q	1	S	A
22		Α	R	E	С	Р	R	V	Р	A
23		A	R	E	С	Р	R	V	S	S
24		Α	R	E	C	P.	R	V	S	Α
25		Α	R	Е	С	Р	R	1	Р	S
26	1	A	R	Е	С	Р	R	Ī	Р	Α
27	 	A	R	E	С	P	R	I	S	S
28	 	A	R	E	Č	P	R	T	·s	Α
	 	A	R	E	R	S	à	V	P	A
29	 		R	. E	R	S	à	V	s	S
30	 	A		E	R	S	Q	V	s	A
31	<u> </u>	A	R						P	S
32		Α	R	E	R	S	Q			
33	<u> </u>	Α	R	E	R	S	Q	<u> </u>	Р	A
34		Α	R	E	R	S	Q	<u> </u>	S	S
35		Α	R	E	R	S	Q		S	Α
36	1	Α	R	E	R	S	R	V	Р	A
37	1	Α	R	E	R	S	R	V	S	S
38		A	R	E	R	S	R	V	S	Α
39	1 1	A	R	E	R	S	R		Р	S
40	i	A	R	E	R	S	R		Р	A
41	 	A	R	Ē	R	s	R	ī	S	S
42	 	A	R	Ē	R	s	R	i	S	Ā
	1 - 1		R	E	R	P	Q	l v	P	A
43	 	A			R	P	Q	V	S	S
44	 !	A	R	E				V	S	A
45	<u> </u>	A	R	E	R	Р	Q	· ·		
46		Α	R	E	R	Р	Q	 	Р	S
47		Α	R	E	R	Р	Q		Р	A
48	1	Α	R	E	R	Р	Q		S	S
49		A	R	E	R	Р	Q		S	Α
50	1 	A	R	Ė	R	Р	R	V	Р	Α
51	 	A	R	E	R	Р	R	V	S	S
52	 	A	R	E	R	P	R	V	s	A
53	 	A	R	E	R	P	R	i	P	S

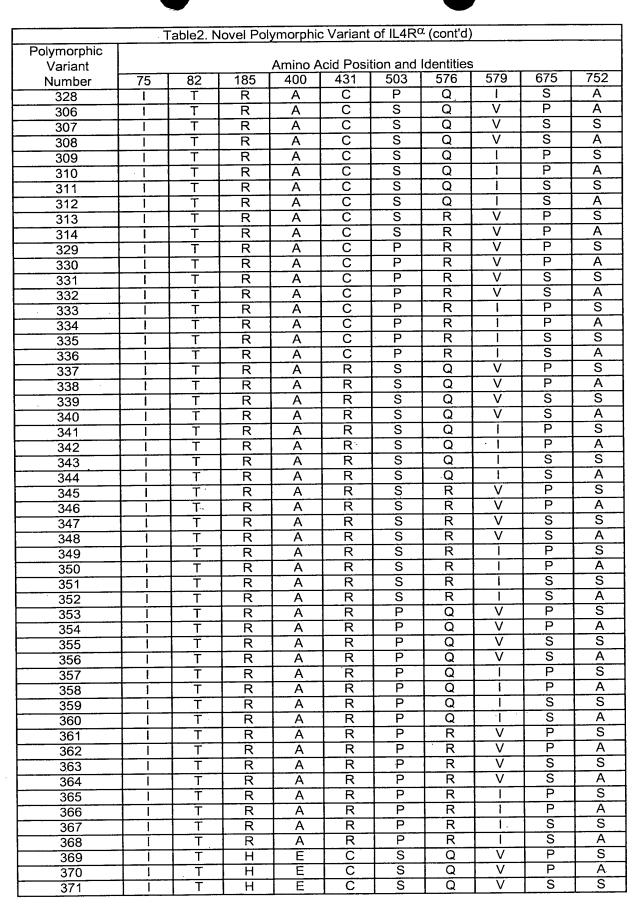
	T:	able2. N	ovel Pol	morphic	Variant	of IL4Rα	(cont'd)			
Polymorphic										
Variant				Amino A	cid Posit	ion and I	dentities			
Number	75	82	185	400	431	503	576	579	675	752
54	1	Α	R	E	R	Р	R	<u> </u>	Р	A
55		A	R	E	R	Р	R	. 1	S	S
56	-	A	R	E	R	Р	R	1	S	Α
57	i	A	R	Α	С	S	Q	V	Р	Α
58		A	R	Α	С	S	Q	V	S	S
.59	- i - l	A	R	A	С	S	Q	V	S	Α
60		Â	R	A	С	S	Q		Р	S
61		A	R	A	С	S	Q		Р	Α
62	- 	A	R	A	č	S	Q	- T	S	S
		$\frac{2}{A}$	R	Â	$\frac{\tilde{c}}{c}$	s	Q	i i	S	A
63	- ; 	$\frac{2}{A}$	R	A	Č	S	R	$\overline{}$	Р	Α
64		$\frac{2}{A}$	R	A	Č	s	R	V	S	S
65		A	R	Ā	C	S	R	v	s	A
66	1.		R	A	C	S	R	i	P	S
67		A			C	S	R		P	A
68		A	R	A	C	S	R		S	S
69		Α	R	A	C	S	R	<u> </u>	S	$\frac{3}{A}$
70	_	A	R	A	C	P	Q	V	P	$\frac{\hat{A}}{A}$
71		Α	R	A				- ×	S	S
- 72		A	R	Α	С	Р	α	V	S	Ā
73	1	A	R	Α	С	Р	Q		P	s
74		Α	R	Α	С	Р	. Q	<u> </u>		
75	1	Α	R	Α	С	Р	Q	1	Р	Α
76	l	Α	R	Α	С	Р	Q		S	S
77	Ī ··	Α	R	.A	С	Р	Q_		S	Α_
78	1	Α	R	A	С	Р	R	V	Р	A
79	. 1	Α	R	Α	С	Р	R	V	S	S
80	1	Α	R	Α	С	Р	R	V	S	A
81		A	R	Α	С	Р	R	l	Р	S
82		Α	R	Α	С	P	R		Р	Α
83		Α	R	Α	С	Р	R		S	S
84		Α	R	Α	C	Р	R		S	Α_
85	 	Α	R	Α	R	S	Q		Р	Α
86		A	R	Α	R	S	Q	V	S	S
87	 	A	R	Α	R	S	Q	V	S	Α
88	l i	A	R	Α	R	s	Q	1	Р	S
89	i	A	R	. A	R	S	Q	T I	Р	Α
90	 	A	R	A	R	S	Q	ı	S	S
91	 	A	R	A	R	S	Q	T	S	Α
92	 	Â	R	A	R	S	R	V	Р	Α
93	 	A	R	A	R	s	R	V	S	S
93	+	A	R	A	R	s	R	V	s	A
	+	A	R	A	R	s	R	1 1	P	S
95	+	A	R	A	R	S	R	l i	P	A
96			R	A	R	S	R	+i	s	S
97	 	A			R	S	R	t i	s	A
98		A	R	A	R	P	à	l v	P	A
99	<u> </u>	A	R	A		P	l a	l v	S	s
100	 	A	R	A_	R		1 a	 v	s	A
101		A	R	A	R	P			P	S
102		A	R	A	R	P	Q		P	A
103	!	A	R	A	R	P	Q			S
104		Α	R	A	R	P	Q_	 	S	
105	1	Α	R	Α	R	Р	Q		S	A
106		Α	R	Α	R	Р	R	V	Р	Α

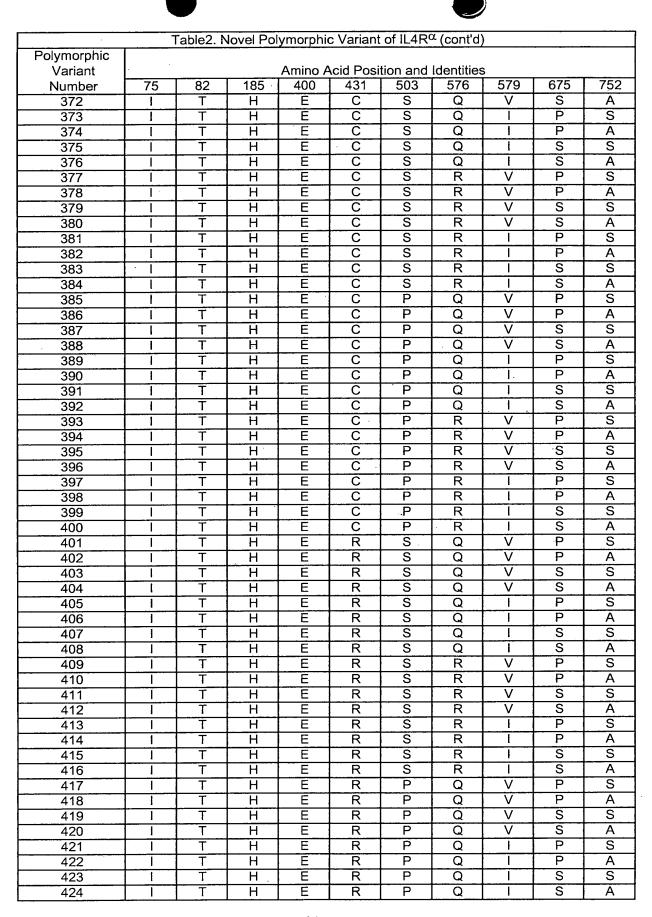
	T	able2. N	ovel Poly	morphic	Variant	of IL4R ^o	(cont'd)			· ·
Polymorphic				Amino A					,	
Variant	75	82	185	400	431	503	576	579	675	752
Number	75	A A	R R	A	R	P	. R	V	s	S
107	'_	$\frac{A}{A}$	R	Â	R	P	R	V	s	A
108			R	Â	R	P	R	i -	P	S
109	l	A	R	Â	R	P	R	- 	P	Ā
110	l l	$\frac{A}{A}$	R	A	R	P	R	 ; -	s	S
111	1		R	Â	R	P	R	<u> </u>	s	A
112	!	A	H	Ē	·c	S	Q	·	P	S
113	1	A		- - -	- C	S	Q	v	P	A
114		A	Н		- C	S	Q	V	s	S
115	1	A	H	E	· C	S	Q	V	S	<u>_</u>
116		Α	H	E	C	S	α	V	P	s
117		Α	H	E		S	Q	<u>-</u>	P	
118	<u> </u>	Α	Н	E	С				S	S
119		Α	Н	E	С	S	Q		S	
120	l	Α	Н	E	C	S	Q	V) P	A S
121		Α	H	E	C	S	R			
122	l	Α	Н	E	C	S	R	>	P	<u>A</u>
123	i	Α	Н	Е	C	S	R	V	S	S
124		Α	Н	E	O	S	R	V	S	A
125		Α	Н	E	C	S	R		P	S
126	1	Α	Ι	E	С	S	R	<u> </u>	Р	Α
127		Α	Η	E	С	S	R		S	S
128	1	Α	Η	E	С	S	R		S	Α
129	I	Α	Н	E	O	Р	Q	V	Р	S
130	I	Α	Н	Е	C	Р	Q	V	Р	A
131	ı	Α	Н	E	С	Р	Q	V	S	S
132	T	Α	Н	E	С	Р	Q	V	S	Α
133	1	Α	Н	E	С	Р	Q	1	P	S
134		• А	Н	E	С	Р	Q	1	Р	Α
135		Α	Н	Е	С	Р	Q		S	S
136	 	Α	Н	Е	С	Р	Q	1	S	Α
137	 	A	Н	E	С	Р	R	V	Р	S
138	i	A	Н	E	С	Р	R	V	P.	Α
139		A	Н	Е	С	Р	R	V	S	S
140	 	A	H	E	С	Р	R	V	S	А
141	 	A	Н	E	С	Р	R		Р	S
142	 	A	H	Ē	Ċ	P	R		Р	Α
143	 	A	H	E	č	P	R	T	s	S
144	 	A	H	Ē	Č	P	R	ı	S	A
145	+ ;	Â	H	E	R	s	Q	V	P	S
146	1	Â	H	E	R	s	à	T V	Р	Α
147	1 1	A	H	Ē	R	S	à	V	S	S
147	 -	A	H	E	R	s	Q	l v	Š	A
	 	A	H	E	R	s	Q	i	P	S
149	1 1	A	H	E	R	s	à	l i	P	A
150			H	E	R	S	l a	 	S	S
151	1 !-	A		 		S	Q	 	s	A
152	 	A	Н		R	S	R	 	P	s
153	!	A	H	E	R			 	P	A
154	1 !	A	H	E	R	S	R	 	S	S
155		A	Н	E	R	S	R			
156		Α	H	E	R	S	R	V	S	A
157		Α	H	E	R	S	R		P	S
158		Α	Н	E	R	S	R	<u> </u>	P	A
159	1	Α	Н	E	R	S	R		S	S

				 		6 II 4 D O	((11-1)			···
		able2. N	ovel Pol	ymorphic	: Variant	of IL4R	(conta)			
Polymorphic				Λmino Λ	oid Doci	tion and	Idantitias			
Variant	75	82	185	400	431	503	576	579	675	752
Number	75			#00 E	R	303 S	R	3/3	S	A
160	- !	A	<u>H</u>	E	R	P	à	- 	P	S
161	<u> </u>	A	H	E	R	P	a	- v	P	A
162		Α	H			P	Q	-v	S	$\frac{2}{s}$
163		A	Н	Ш	R	P		V	S	<u>S</u>
164		Α	Н	E	R		Q		P	_ <u>^</u>
165	1	Α	H	E	R	Р	Q			
166		Α	H	E	R	Р	Q	1	Р	A
167	1	Α	Н	Е	R	Р	Q		S	S
168	<u> </u>	Α	H	E	R	P	α		S	A
169		Α	Η	E	R	Р	R	V	Р	S
170		Α	H	E	R	Р	R	V	Р	A
171		Α	Η	E	R	P	R	V	S	S
172	L	Α	Τ	E	R	Р	R	V	S	A
173	ı	Α	H	E	R	Р	R	1	Р	S
174	Î	Α	Ι	E	R	Р	R		Р	Α
175	l l	. A	Н	Е	R	Р	R	l l	S	S
176		Α	Н	Е	R	Р	R	1	S	Α
177		Α	Н	Α	С	S	Q	V	Р	S
178	· 1	Α	Н	Α	С	S	Q	٧	Р	Α
179	ī	Α	Н	Α	С	S	Q	٧	S	S
180		Α	Н	Α	С	S	Q	V	S	Α
181	ti	Α	Н	Α	С	S	α		Р	S
182	i	A	Н	Α	С	S	Q	l i	Р	Α
183	l i	A	Н	Α	С	S	Q	1	S	S
184	 	A	H	A	С	S	Q	1	S	Α
185	 	A	H	A	Ċ	S	R	V	Р	S
186	 	A	H	A	Ċ	S	R	V	Р	Α
187	 	A	H	Ā	Ċ.	s	R	V	S	S
188	 	A	H	A	Č	s	R	V	S	Α
189	 	A	H	A	c	s	R		P	S
190	 	A	H	A	C	s	R	Hi	Р	Α
191	1 1	Â	Н	A	C	Š	R	l i	S	S
192	<u> </u>	A	Н	A	C	s	R	i i	s	A
	 	 A	H	A	C	P	Q	V	P	S
193	 	A	H	A	C	P	à	V	P	A
194 195	 	A	H	A	C	P	à	l v	s	S
	 	A	H	A	C	P	Q	l v	s	Ā
196		A	H	A	C	P	Q	ì	P	S
197	 	A	H	Â	C	P	Q	 	P	A
198	+ +		H	A	c	P	Q	 	s	s
199	1	A	H	A	C	P	a	 	s	Ā
200	<u> </u>	A	H		 c	P	R	 '	P	s
201	<u> </u>	A		A	c	P	R	l v	P	A
202		A	H	A	C	P	R	V	s	S
203		A	H	A	C	P	R	l v	s	A
204	1 -	A			C	P	R	 	P	S
205	+	A	H	A	C	P	R	 	P	A
206	 	A	H	A		1		+	S	S
207	1 !	A	Н	A	C	P	R	<u> </u>		
208	<u> </u>	A	Н	A	C	P	R	1 7	S	S
209		A	H	A	R	S	l Q	V	P	
210	1 1	Α	Н	A	R	S	Q_	V	Р	A
211		A	H	A	R	S	Q	V	S	S
212		Α	H	A	R	S	Q	V	S	A

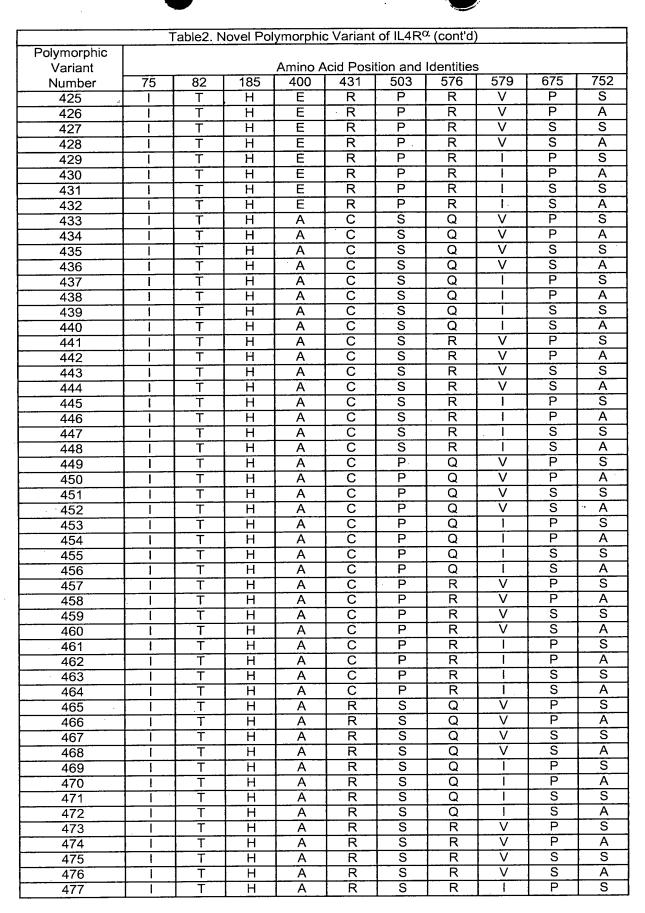
		able2. N	ovel Pol	ymorphic	: Variant	of IL4R ^o	(cont'd)			
Polymorphic										
Variant				Amino A					C75	750
Number	75	82	185	400	431	503	576	579	675 P	752 S
213	!	<u> </u>	H	A	R	S	<u>Q</u>	1	P	A
214		A	<u>H</u>	Α	R	S	Q		S	S S
215	l l	<u> </u>	Н	Α	R	S	Q		S	
216		Α	Н	Α	R	S	<u>Q.</u>	V	P	A S
217		Α	H	Α	R	S	R	V	P	A
218	!	Α	H	A	R	S	R R	V	S	S
219		A	Н	A	R	S	R	V	S	A
220	_ !	A	H	Α	R	S	R	V 1	P	S
221		Α	H	A	R	S	R	1	P	A
222		Α	Н	Α	R	S	R	<u> </u>	S	S
223	<u>l</u>	Α	H	A	R	S	R		S	A
224	1	A	H	A	R	P	Q	V	- 3 P	S
225	- !	Α	H	A	R	P		V	P	A
226	1	A	H	A	R	P	Q	V	S	S
227		A	Н	A	R R	P	Q	V	S	A
228		Α	Н	A		P	<u>3</u> Q	Ī	P	S
229		A	H	A	R R	P	Q	1	P	A
230	1	A	H	A	R	P	α		S	s
231	1	A	Н	A	R	P	a	<u> </u>	S	Ā
232	<u> </u>	A	H	A	R	P	R	- '-	P	S
233	<u> </u>	A	H		R	P	R	V	P	A
234	<u> </u>	A	H	A	R	P	R	l v	S	s
235	<u> </u>	A			R	P	R	V	S	A
236	l	A	Н	A	R	P	R	- `	P	S
237	1	A	H	A	R	P	R	 	P	Ā
238		A	H	A	R	P	R		s	S
239	1	A	H	A	R	P	R	l-i-	·s	Ā
240		A T	R	Ê	C	s	à	l v	P	S
241 242	<u> </u>	+	R	E	C	S	Q	V	P	Ā
	1	 	R	E	C	S	Q	l v	s	S
243 244		 	R	E	C	s	ā	v	s	Ā
244	1 1	 	R	E	C	S	Q	1	P	s
245	 	 	R	E	C	Š	à	i	P	A
247	 	+	R	E	C	s	à	 	S	s
	 	 	R	T E	Ċ	Š	à	1	S	Α
248 249	1.	╀╌┼┈	R	Ē	C	S	R	V	P	S
250	 	 	R	Ē	C	s	R	V	P	A
251	 	 	R	E	C	Š	R	V	S	S
252	 	 	R	E	c	S	R	V	S	Α
253	l i	Ť	R	 E	Ċ	s	R	1	Р	S
254	i	T	R	E	c	S	R	1	Р	Α
255	 	Ť	R	E	C	S	R	l i	S	S
256	i	Ť	R	T E	c	S	R	1	S	Α
257	 	 	R	E	C	Р	Q	V	Р	S
258	l i	 	R	E	С	Р	Q	V	Р	Α
259	l i	 	R	E	С	Р	Q	V	S	S
260	1 i	i i	R	E	С	Р	Q	V	S	Α
261	 	† †	R	E	С	Р	Q		Р	S
262	l i	 	R	E	С	Р	Q	1	Р	Α
263	l i	 	R	E	c	Р	Q	1	S	S
264	l i	 	R	TE	Ċ	Р	Q		S	Α
265	t i	 	R	E	C	P	R	V	Р	S

		Table2. N	lovel Po	lymorphi	o Variant	of II ADO	½ (cont'd)			
Polymorphic		rablez. N	overro	утпогрти	C Variani	OI IL-41	(COITE O	<u>'</u>	· · · ·	· · ·
				Amino /	\cid Doci	tion and	Identities			
Variant	75	82	185	400	431	503	576	579	675	752
Number		02 T		E E	C 431	903 P	776 R	V V	P	
266			R					_		A S
267	<u> </u>	T	R	E	C	Р	R	V	S	
268	1	Т	R	E	С	Р	R	V	S	<u>A</u>
269	I	T	R	E	С	Р	R		Р	S
270	l	T	R	E	С	Р	R	l l	Р	A
271	l	T	R	E	C	Р	R		S	S
272	1	Ŧ	R	E	С	Р	R	l	S	Α
273	i i	Т	R	E	R	S	Q	V	Р	S
274	1	T	R	E	R	S	Q	V	Р	A
275		Т	R	Е	R	S	Q	V	S	S
276	i	Т	R	E	R	S	Q	V	S	Α
277		Ť	R	Ē	R	s	Q	ı	Р	S
278	<u> </u>	Ť	R	. E	R	s	à	i	P	Ā
279		T	R	E	R	S	Q		S	S
			R	E	R	S	Q	1	S	A
280	<u> </u>	T		E	R	S	R	V	P	- S
281	<u> </u>	T	R					V	P	
282	<u> </u>	T	R	E	R	S	R			A
283	!	T	R	E	R	S	R	> ;	S	S
284		Т	R	E	R	S	R	V	S	Α
285		T	R	Ε	R	S	R	l	Р	S
286	_	T	R	Ε	R	S	R		Р	Α
287	ì	T	R	Е	R	S	R	ı	S	S
288	1	T	R	E	R	S	R	ı	S	Α
289	1	T	R	E	R	Р	Q	V	Р	S
290	ı	T	R	E	R	Р	Q	V	Р	. A
291	i	Т	R	E	R	Р	Q	V	S	Ś
292	- i	i i	R	Ē	R	P	Q	V	S	A
293	i	Ť	R	Ē	R	P	Q	i	P	S
293		T	R.	E	R	P	Q	<u> </u>	P	A
295	1	 '	R	E	R	P	ä	<u>'</u>	s	S
		 		늗늗	R	P	a		S	A
296		1	R	<u> </u>	1	P		- ' -	P	S
297	1	T	R	E	R		R			
298		T	R	E	R	Р	R	V	P	A
299		T	R	E	R	Р	R	V	S	S
300	- 1	T	R	E	R	Р	R		S	Α
301	l	T	R	E	R	Р	R	1	Р	S
302	ı	T	R	E	R	Р	R	1	Р	Α
303	-	Т	R	E	R	Р	R	1	S	S
304	ı	Ť	R	Е	R	P	R	I	S	Α
305	ı	Т	R	Α	С	S	Q	V	Р	S
315	1	Ť	R	Α	C	s	R	V	S	S
316	i	 	R	A	Ċ	s	R	V	S	A
317	i	 	R	A	Č	Š	R	i i	P	S
318	<u> </u>	 	R	A	Č	s	R	i	P	A
319	<u> </u>	T	R	A	C	S	R	- 	S	S
	1	1			č	S	R		S	A
320		T	R	A				\ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \ \	P	s
321	<u> </u>	T	R	A	С	Р	Q			
322		T	R	A	С	Р	Q	V	Р	A
323		T	R	A	С	Р	Q	V	S	S
324		T	R	Α	С	Р	Q	V	S	Α
325	Ī	Т	R	Α	С	P	Q	1	Р	S
326	1 .	Т	R	Α	С	Р	Q		′ P	Α
327	1	T	R	Α	С	Р	Q		S	S

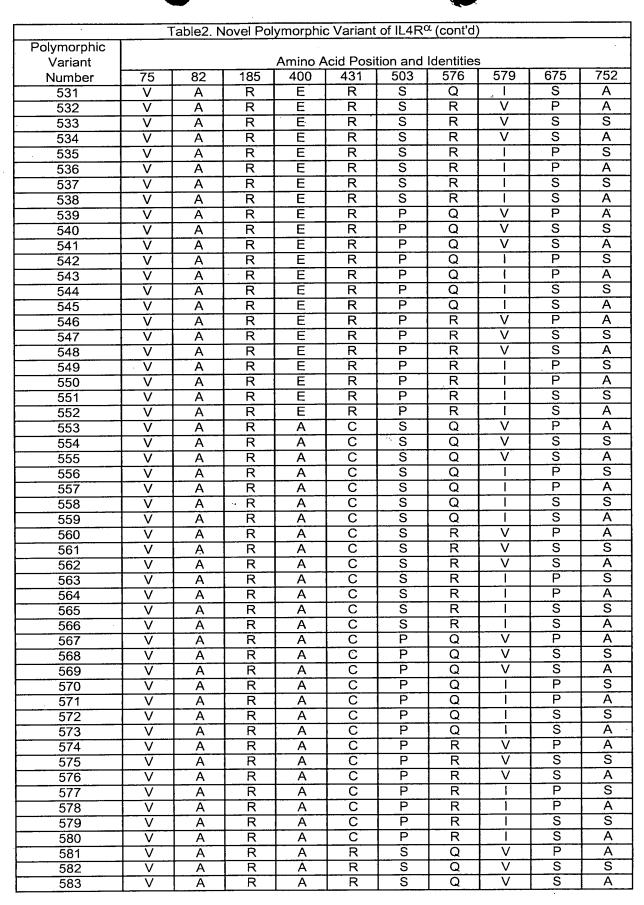




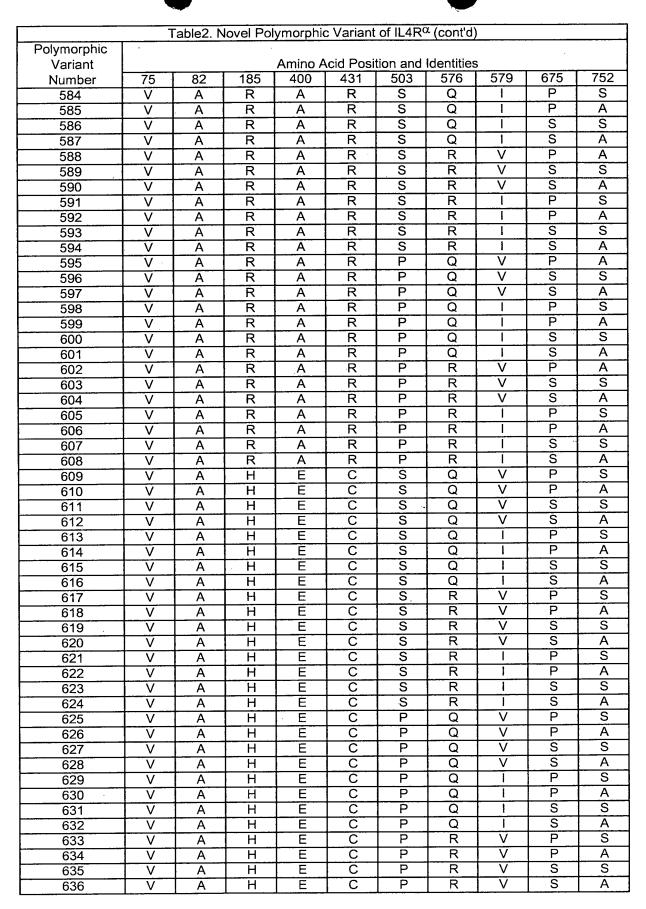
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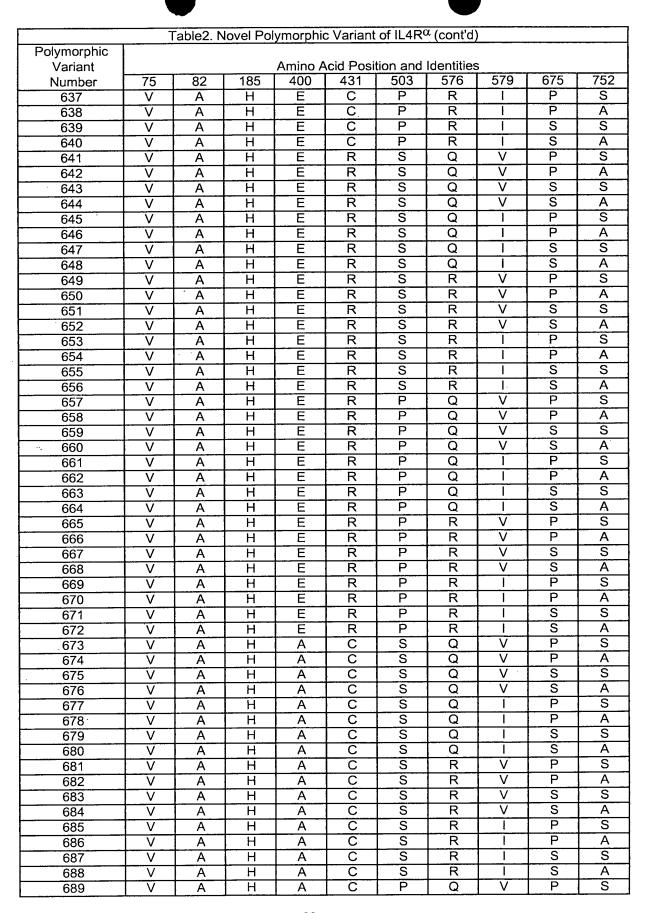


	π	able2. N	lovel Pol	vmorphi	c Variant	of IL4R ^o	^x (cont'd)			
Polymorphic				,						
Variant				Amino A	cid Posi	tion and	Identities	S		
Number	75	82	185	400	431	503	576	579	675	752
478	l	. T	Н	Α.	R	S	R	i	Р	Α
479	1	Т	Н	Α	R	S	R	1	S	S
480	ı	Т	Н	Α	R	S	R	1.	S	Α
481	1	T	Н	Α	R	Р	Q	V	Р	S
482	ı	Т	Н	Α	R	Р	Q	V	Р	Α
483	1	Т	Н	Α	R	Р	Q	V	S	S
484		T	Н	Α	R	Р	Q	V	S	Α
485		Ŧ	Н	A	R	Р	Q		Р	S
486		Т	H	Α	R	Р	Q		Р	Α
487	i i	T	H	Α	·R	Р	Q		· S	S
488	1	Ť	Н	Α	R	Р	Q		S	Α
489	i	Т	Н	Α	R	Р	R	V	Р	S
490		T	Н	• A	R	Р	R	V .	Р	Α
491	i	Ť	Н	A	R	Р	R	V	S	S
492		Ť	H	A	R	Р	R	V	S	Α
493	<u> </u>	T	H	Α	R	Р	R		Р	S
494	i	Ť	H	A	R	Р	R		Р	Α
495	i	T	H	A	R	Р	R	ı	S	S
496		T	Н	Α	R	Р	R		S	Α
497	V	A	R	Е	С	s	Q	V	Р	Α
498	ν	A	R	Ē	Ċ	S	Q	V	S	S
499	⊢ÿ	A	R	Ē	č	S	Q	V	S	A
500	V	A	R	Ē	C	S	à	l	Р	S
501	V	A	R	E	Ċ	S	Q	i	Р	Α
502	l v	A	R	Ē	C	s	Q	i	S	S
503	l v	A	R	Ē	C	s	Q	i	S	Ā
504	V	A	R	Ē	Č	Š	R	V	P	A
505	V	A	R	E	č	Š	R	V	S	S
506	l v	A	R	Ē	Č	s	R	V	S	A
507	V	A	R	Ē	Č	s	R	i	P	S
508	T V	Ā	R	Ē	c	s	R	i i	Р	A
509	V	A	R	Ē	Č	s	R		S	S
510	V	A	R	Ē	C	s	R	i	S	A
511	l v	A	R	Ē	C	P	Q	V	P	A
512	l v	Ā	R	È	C	P	à	V	S	S
513	l v	A	R	E	C	P	Q	V	s	A
514	V	A	R	E	C	P	Q	1	P	S
515	V	A	R	Ē	C	P	à	i	P	A
516	l v	A	R	Ē	C	P	Q	i	S	S
517	V	A	R	E	C	P	à	i	S	Ā
518	V	Â	R	E	C	P	R	V	P	A
519	V	Â	R	Ē	C	P	R	V	s	S
520	 	A	R	E	C	P	R	V	s	Ā
520	V	Â	R	E	c	P	R	i	P	s
521	V	A	R	E	C	P	R		P	Ā
523	V	A	R	E	C	P	R	i	S	S
524	l v	Â	R	Ē	C	P	R	i	s	Ā
525	l v	A	R	눝	R	s	Q	V	P	A
526	l v	A	R	E	R	s	à	V	S	s
527	 	A	R	Ė	R	s	Q	V	S	A
527	V	A	R	E	R	s	Q	 '	P	s
	 v	A	R	E	R	S	à	'	P	A
529 530	V	A	R	E	R	S	Q	 	s	S
530				<u> </u>			<u> </u>	L'		



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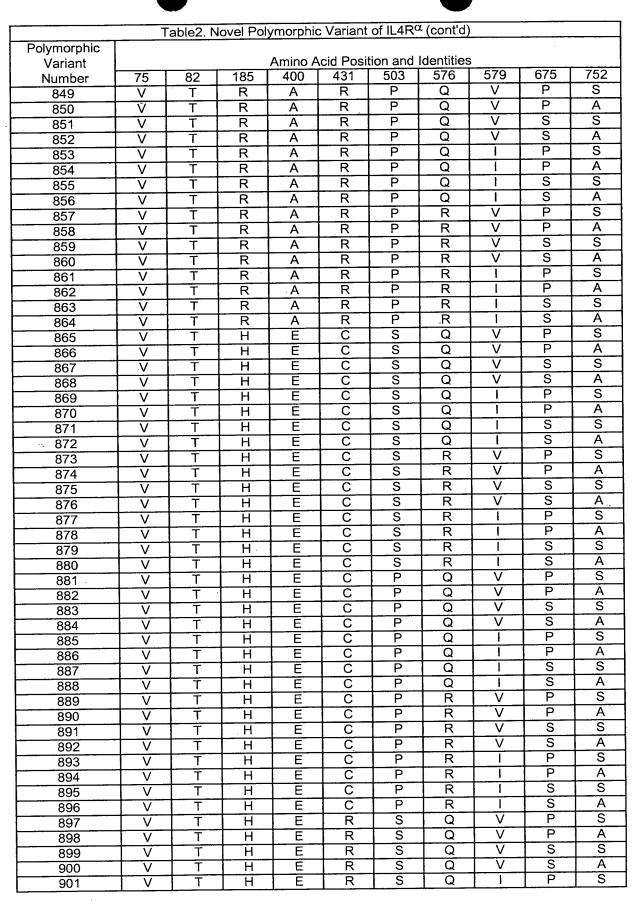


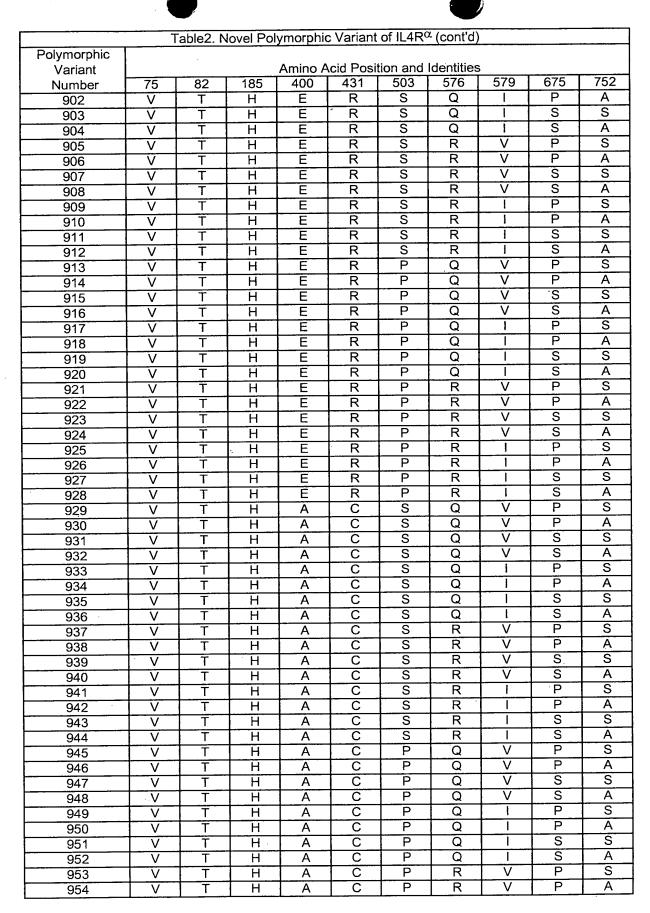


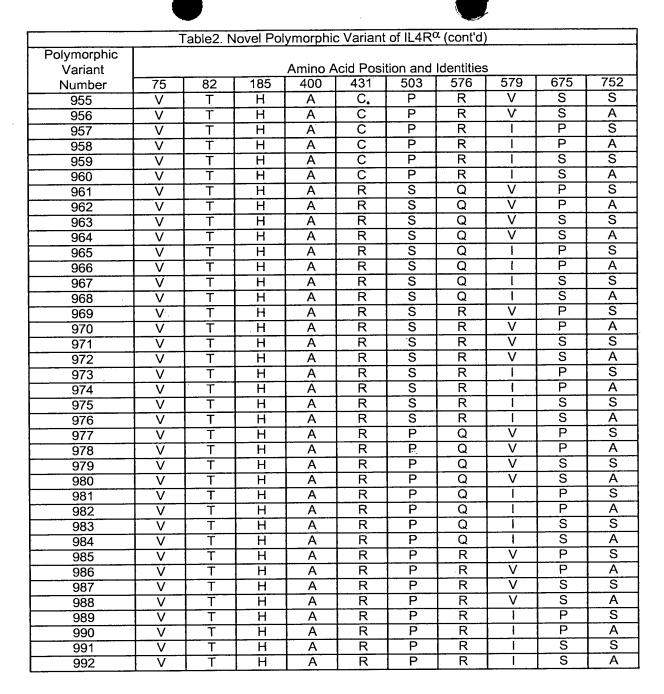
	т	able2 N	ovel Pol	ymorphic	Variant	of IL4R ^c	(cont'd)			
Polymorphic		abicz. IN	OVEI 1 OI	ymor princ	variant	OI IL II V	(oonea)			-
Variant				Amino A	cid Posit	ion and	Identities			
Number	75	82	185	400	431	503	576	579	675	752
690	V	A	H	A	C	P	Q	V	Р	Α
691	V	$\frac{\Lambda}{A}$	H	A	C	P	ã	V	S	S
692	l v	A	H	A	C	P	Q	V	s	Ā
693	V	Â		A	C	P	Q	i	P	S
	V	$\frac{A}{A}$	H	A	C	P	Q	<u> </u>	P	A
694	V	A	H	A	C	P	Q	<u>_</u>	S	S
695	V	A	Н	A	C	P	Q	<u>'</u>	s	Ā
696			Н	A	C	Р	R	V	P	s
697	V	A		A	C	P	R	V	Р	A
698	V	A	H		C	P	R	V	S	s
699	V	A	H	A				V	S	A
700	V	Α	H	A	υ	Р	R	<u>V</u>	P	S
701	V	Α	H	A	υ	Р	R	<u> </u>		
702	V	A	H	Α	υ	P 2	R	1	PS	A_S
703	V	Α	H	A	υ	Р	R	<u> </u>		
704	V	Α	Н	Α	С	Р	R	1	S	A S
705	V	Α	Н	Α	R	S	Q	V	Р	
706	V	Α	Н	Α	R	S	Q	V	Р	A
707	V	Α	Н	Α	R	S	Q	V	S	S
708	V	Α	Н	Α	R	S	α	V	S	A
709	V	Α	H	Α	R	S	Q		Р	S
710	V	Α	H	Α	R	S	Q		Р	<u>A</u>
711	V	Α	H	Α	R	S	Q		S	S
712	V	Α	Н	Α	R	S	Q	<u> </u>	S	Α
713	V	Α	Н	Α	R	S	R	V	Р	S
714	V	Α	Н	Α	R	S	R	>	P	Α
715	V	Α	Н	Α	R	S	R	V	S	S
716	V	Α	Н	Α	R	S	R	V	S	Α
717	V	A	Н	Α	R	S	R		Р	S
718	V	Α	Н	Α	R	S	R	I	P	Α
719	$\vdash \lor$	Α	Н	A	R	S	R	I	S	S
720	V	Α	Н	Α	R	S	R	1	S	Α
721	V	Α	Н	A	R	Р	Q	V	Р	S
722	T V	Α	Н	Α	R	Р	Q	·V	Р	Α
723	l v	A	Н	Α	R	Р	Q	V	S	S
724	V	Α	Н	A	R	Р	Q	V	S	· A
725	İ	A	H	A	R	Р	Q	i i	. P	S
726	l v	A	H	A	R	Р	Q	Ī	Р	Α
727	l v	A	H	A	R	P	Q	1	S	S
728	l v	A	H	A	R	P	Q	Ī	S	Α
729	l v	A	H	A	R	P	R	V	Р	S
730	l v	A	H	A	R	P	R	V	Р	A
731	l v	Â	H	A	R	P	R	l v	S	S
732	l v	A	H	A	R	P	R	 v	s	A
732	V	A	H	A	R	P	R	l i	P	S
734	<u>v</u>	A	H -	A	R	P	R	 	P	Ä
735	 	 	H	A	R	P	R	 	s	S
	V		H	A	R	P	R	 	S	A
736		A T		E	C	S	Q	 '	P	s
737	V		R		C	S	Q	 	P	A
738	V	T	R	E				 	S	S
739	V	T	R	E	C	S	Q	 	S	A
740	V	T	R	E	C	S	Q		P	
741	V	<u> </u>	R	E	C	S	Q			S
742	V	T	R	E	С	S	Q		Р	A

						5 II 4 DO				
		able2. N	ovel Pol	ymorphic	: Variant	of IL4R ^o	(contid)			
Polymorphic					:	السياسية				
Variant		- 00 1				ion and	576	579	675	752
Number	75	82	185	400	431	503			8 S	75 <u>2</u> S
743	V	Ţ	R	E	С	S	Q	1 -	S	A
744	. V	T	R	E	С	S	Q		P	S
745	V	T	R	E	С	S	R	V		
746	V	Т	R	E	C	S	R	V	Р	A
747	V	Т	R	E	O	S	R	V	S	S
748	V	T	R	Ε	O	S	R	V	S	A
749	V	T	R	E	C	S	R		Р	S
750	V	Ť	R	E	С	S	R	ı	Р	Α
751	V	Τ	R	E	C	S	R		S	S
752	V	Т	R	Ε	O	S	R		S	Α
753	V	Т	R	E	С	Р	Q	V	Р	S
754	V	Т	R	E	С	Р	α	V	P	Α
755	V	Т	R	Е	C	Р	Q		S	S
756	V	T	R	E	C	Р	Q	V	S	Α
757	V	Ť	R	Е	С	Р	α	ı	Р	S
758	V	T	R	Ĕ	С	Р	Q		Р	Α
759	V	T	R	Е	С	Р	α		S	S
760	l i	Ť	R	Е	С	Р	Q	Ī	S	Α
761	l v	Ť	R	E	С	Р	R	V	Р	S
762	l v	Ť	R	Ē	Ċ	Р	R	V	Р	Α
763	l v	T	R	E	Č	P	R	V	S	S
764	l v	Ť	R	E	c	Р	R	V	S	Α
765	l v l	- 	R	E	Č	P	R	i	Р	S
765	l v	T	R	Ē	C	Ρ	R	i	P	Ā
767	 	+	R	E	C	P	R	i i	S	S
	V	<u>'</u>	R	E	C	P	R	i	S	Ā
768	V	- '-	R	E	R	s	Q	V	P	S
769	 v	+	R.	E	R	S	à	V	P	Ā
770	I		1	E	R	S	Q Q	V	s	S
771	V	T	R		R	S	à	⊢ v ⊤	S	A
772	V	T	R	E				V	P	s
773	V	T	R	E	R	S	Q	<u> </u>	P	A
774	V	T	R	E	R	S	Q	 	S	S
775	V	<u> </u>	R	E	R	S	Q		S	
776	V	T	R	E	R	S	Q	<u> </u>	P	A
777	V	T	R	E	R	S	R	V		S
778	V	T	R	Е	R	S	R	V	P	A
779	V	Ť	R	E	R	S	R	V	S	S
780	V	Т	R	E	R	S	R	<u> </u>	S	A
781	V	T	R	E	R	S	R	<u> </u>	P	S
782	V	Ť_	R	E	R	S	R	<u> </u>	Р	A
783	V	T	R	E	R	S	R		S	S
784	V	Т	R	Ε	R	S	R		S	A
785	V	T	R	E	R	Р	Q	V	Р	S
786	V	Т	R	Е	R	Р	Q	V	Р	Α_
787	V	Т	R	Е	R	Р	Q	V	S	S
788	V	T	R	Е	R	Р	Q	V	S	Α
789	V	Т	R	E	R	Р	Q		Р	S
790	T V	T	R	Е	R	Р	Q		Р	Α
791	l v	T	R	E	R	Р	Q		S	S
792	 v -	Ť	R	T E	R	Р	Q		s	Α
793	l v	T	R	E	R	P	R	V -	P	S
794	$+\dot{\overline{}}$	 	R	F	R	P	R	l v	Р	Ā
795	 	T	R	E	R	P	R	l v	s	S
195		<u> </u>	<u> </u>			<u> </u>	<u> </u>	<u> </u>		

	Т	able2. N	lovel Pol	ymorphic	Variant	of IL4R ^c	^z (cont'd)			
Polymorphic										
Variant							Identities			
Number	75	82_	185	400	431	503	576_	579	675	752
796	٧	Τ	R	E	·R	Р	R	V	S	A
797	V	T	R	E	R	Р	R	<u>l</u>	Р	S
798	V	T	R	Е	R	Р	R	l	Р	Α
799	٧	T	R	E	R	Р	R	ı	S	S
800	V	T	R	E	R	Р	R	l	S	Α
801	V	Τ	R	Α	С	S	Q	V	Р	S
802	V	T	R	Α	С	S	Q	V	Р	A
803	V	Т	R	Α	С	S	Q	V	S	S
804	V	T	R	Α	С	S	Q	V	S	Α
805	V	Т	R	Α	С	S	Q	1	Р	S
806	V	Τ	R	Α	С	S	Q	l	Р	Α
807	V	Т	R	Α	С	S	Q	1	. S	S
808	V	T	R	Α	С	S	Q		S	Α
809	V	T	R	Α	. C	S	R	V	Р	S
810	V	T	R	Α	С	S	R	V	Р	Α
811	V	Т	R	Α	С	S	R	V	S	S
812	V	T	R	Α	С	S	R	V	S	A
813	V	Ť	R	Α	С	S	R	Ī	Р	S
814	V	Ť	R	Α	С	S	R	ı	Р	Α
815	V	Ť	R	A	С	S	R	ï	S	S
816	V	Ť	R	Α	C .	S	R		S	Α
817	l v	T	R	Α	С	P	Q	V	Р	S
818	V	Ť	R	Α	Ċ	Р	Q	V	Р	Α
819	V	Ť	R	A	C	Р	Q	V	S	S
820	V	l i	R	A	c	P	Q	V	S	Α
821	V	Ť	R	A	C	P	à		P	S
822	V	T	R	A	Č	P	à	i	Р	A
823	l v	i	R	A	C	P	. Q	l i	s	S
824	l v	Ť	R	A	C	P	à	i	S	A
825	 v -	Ť	R	A	C	P	R	V	P	S
826	V	' ' 	R	A	C	P	R	l v	P	Ā
	l v	T	R	A	C	P	R	l v	s	S
827	\ \ \ \ \	' -	R	Â	C	P	R	V	s	Ä
828	V	 '	R	Â	C.	P	R	1 1	P	S
829	 	+	R	A	C.	P	R	1	P	Ā
830		 		A	 c	P	R	1	S	s
831	V	+	R	A	 c	P	R	' -	s	A
832	V	 	R	A	R	S	Q	l v	P	S
833	V		R		R	S	Q	V	P	Ā
834	V	T	R	A	R	S	Q	l v	S	S
835	V _	T	R	A	R	S	l a	V	S	A
836	V	T	R	A		S	Q	 	P	S
837	V	T	R	A	R	S		-	P	A
838	V	T	R	A	R		Q	'	S	S
839	V	T	R	A	R	S	Q		S	
840	V	Ţ	R	A	R	S	Q	1	P	A
841	V	T	R	A	R	S	R	V	1	S
842	V	I	R	A	R	S	R	V	Р	A
843	V	Т	R	Α	R	S	R	V	S	S
844	V	Т	R	Α	R	S	R	V	S	A
845	V	Τ	R	Α	R	S	R		Р	S
846	V	T	R	Α	R	S	R		Р	A
847	V	T	R	Α	R	S	R		S	S
848	V	Т	R	Α	R	S	R	1	S	Α







The invention also includes IL4R α peptide variants, which are any fragments of an IL4R α protein variant that contains one or more of the amino acid variations shown in Table 2. An IL4R α peptide variant is at least 6 amino acids in length and is preferably any number between 6 and 30 amino acids long, more preferably between 10 and 25, and most preferably between 15 and 20 amino acids long. Such IL4R α peptide variants may be useful as antigens to generate antibodies specific for one of the above IL4R α isoforms. In addition, the IL4R α peptide variants may be useful in drug screening assays.

An IL4R α variant protein or peptide of the invention may be prepared by chemical synthesis or by expressing one of the variant IL4R α genomic and cDNA sequences as described above.

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Alternatively, the IL4R α protein variant may be isolated from a biological sample of an individual having an IL4R α isogene which encodes the variant protein. Where the sample contains two different IL4R α isoforms (i.e., the individual has different IL4R α isogenes), a particular IL4R α isoform of the invention can be isolated by immunoaffinity chromatography using an antibody which specifically binds to that particular IL4R α isoform but does not bind to the other IL4R α isoform.

The expressed or isolated IL4R α protein may be detected by methods known in the art, including Coomassie blue staining, silver staining, and Western blot analysis using antibodies specific for the isoform of the IL4R α protein as discussed further below. IL4R α variant proteins can be purified by standard protein purification procedures known in the art, including differential precipitation, molecular sieve chromatography, ion-exchange chromatography, isoelectric focusing, gel electrophoresis, affinity and immunoaffinity chromatography and the like. (Ausubel et. al., 1987, In Current Protocols in Molecular Biology John Wiley and Sons, New York, New York). In the case of immunoaffinity chromatography, antibodies specific for a particular polymorphic variant may be used.

A polymorphic variant IL4R α gene of the invention may also be fused in frame with a heterologous sequence to encode a chimeric IL4R α protein. The non-IL4R α portion of the chimeric protein may be recognized by a commercially available antibody. In addition, the chimeric protein may also be engineered to contain a cleavage site located between the IL4R α and non-IL4R α portions so that the IL4R α protein may be cleaved and purified away from the non-IL4R α portion.

An additional embodiment of the invention relates to using a novel IL4R α protein isoform in any of a variety of drug screening assays. Such screening assays may be performed to identify agents that bind specifically to all known IL4R α protein isoforms or to only a subset of one or more of these isoforms. The agents may be from chemical compound libraries, peptide libraries and the like. The IL4R α protein or peptide variant may be free in solution or affixed to a solid support. In one embodiment, high throughput screening of compounds for binding to an IL4R α variant may be accomplished using the method described in PCT application WO84/03565, in which large numbers of test compounds are synthesized on a solid substrate, such as plastic pins or some other surface, contacted with the IL4R α protein(s) of interest and then washed. Bound IL4R α protein(s) are then detected using methods well-known in the art.

In another embodiment, a novel IL4R α protein isoform may be used in assays to measure the binding affinities of one or more candidate drugs targeting the IL4R α protein.

In another embodiment, the invention provides antibodies specific for and immunoreactive with one or more of the novel IL4R α variant proteins described herein. The antibodies may be either monoclonal or polyclonal in origin. The IL4R α protein or peptide variant used to generate the antibodies may be from natural or recombinant sources or produced by chemical synthesis using synthesis techniques known in the art. If the IL4R α protein variant is of insufficient size to be antigenic, it may be conjugated, complexed, or otherwise covalently linked to a carrier molecule to enhance the antigenicity

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of the peptide. Examples of carrier molecules, include, but are not limited to, albumins (e.g., human, bovine, fish, ovine), and keyhole limpet hemocyanin (Basic and Clinical Immunology, 1991, Eds. D.P. Stites, and A.I. Terr, Appleton and Lange, Norwalk Connecticut, San Mateo, California).

In one embodiment, an antibody specifically immunoreactive with one of the novel IL4R α protein isoforms described herein is administered to an individual to neutralize activity of the IL4R α isoform expressed by that individual. The antibody may be formulated as a pharmaceutical composition which includes a pharmaceutically acceptable carrier.

Antibodies specific for and immunoreactive with one of the novel IL4R α protein isoform described herein may be used to immunoprecipitate the IL4R α protein variant from solution as well as react with IL4R α protein isoforms on Western or immunoblots of polyacrylamide gels on membrane supports or substrates. In another preferred embodiment, the antibodies will detect IL4R α protein isoforms in paraffin or frozen tissue sections, or in cells which have been fixed or unfixed and prepared on slides, coverslips, or the like, for use in immunocytochemical, immunohistochemical, and immunofluorescence techniques.

In another embodiment, an antibody specifically immunoreactive with one of the novel IL4R α protein variants described herein is used in immunoassays to detect this variant in biological samples. In this method, an antibody of the present invention is contacted with a biological sample and the formation of a complex between the IL4Ra protein variant and the antibody is detected. As described, suitable immunoassays include radioimmunoassay, Western blot assay, immunofluorescent assay, enzyme linked immunoassay (ELISA), chemiluminescent assay, immunohistochemical assay, immunocytochemical assay, and the like (see, e.g., Principles and Practice of Immunoassay, 1991, Eds. Christopher P. Price and David J. Neoman, Stockton Press, New York, New York; Current Protocols in Molecular Biology, 1987, Eds. Ausubel et al., John Wiley and Sons, New York, New York). Standard techniques known in the art for ELISA are described in Methods in Immunodiagnosis, 2nd Ed., Eds. Rose and Bigazzi, John Wiley and Sons, New York 1980; and Campbell et al., 1984, Methods in Immunology, W.A. Benjamin, Inc.). Such assays may be direct, indirect, competitive, or noncompetitive as described in the art (see, e.g., Principles and Practice of Immunoassay, 1991, Eds. Christopher P. Price and David J. Neoman, Stockton Pres, NY, NY; and Oellirich, M., 1984, J. Clin. Chem. Clin. Biochem., 22:895-904). Proteins may be isolated from test specimens and biological samples by conventional methods, as described in Current Protocols in Molecular Biology, supra.

Exemplary antibody molecules for use in the detection and therapy methods of the present invention are intact immunoglobulin molecules, substantially intact immunoglobulin molecules, or those portions of immunoglobulin molecules that contain the antigen binding site. Polyclonal or monoclonal antibodies may be produced by methods conventionally known in the art (e.g., Kohler and Milstein, 1975, Nature, 256:495-497; Campbell Monoclonal Antibody Technology, the Production and Characterization of Rodent and Human Hybridomas, 1985, In: Laboratory Techniques in Biochemistry

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and Molecular Biology, Eds. Burdon et al., Volume 13, Elsevier Science Publishers, Amsterdam). The antibodies or antigen binding fragments thereof may also be produced by genetic engineering. The technology for expression of both heavy and light chain genes in E. coli is the subject of PCT patent applications, publication number WO 901443, WO 901443 and WO 9014424 and in Huse et al., 1989, Science, 246:1275-1281. The antibodies may also be humanized (e.g., Queen, C. et al. 1989 Proc. Natl. Acad. Sci. 86;10029).

Effect(s) of the polymorphisms identified herein on expression of IL4Rα may be investigated by preparing recombinant cells and/or organisms, preferably recombinant animals, containing a polymorphic variant of the IL4Rα gene. As used herein, "expression" includes but is not limited to one or more of the following: transcription of the gene into precursor mRNA; splicing and other processing of the precursor mRNA to produce mature mRNA; mRNA stability; translation of the mature mRNA into IL4Rα protein (including codon usage and tRNA availability); and glycosylation and/or other modifications of the translation product, if required for proper expression and function.

To prepare a recombinant cell of the invention, the desired IL4R α isogene may be introduced into the cell in a vector such that the isogene remains extrachromosomal. In such a situation, the gene will be expressed by the cell from the extrachromosomal location. In a preferred embodiment, the IL4R α isogene is introduced into a cell in such a way that it recombines with the endogenous IL4R α gene present in the cell. Such recombination requires the occurrence of a double recombination event, thereby resulting in the desired IL4R α gene polymorphism. Vectors for the introduction of genes both for recombination and for extrachromosomal maintenance are known in the art, and any suitable vector or vector construct may be used in the invention. Methods such as electroporation, particle bombardment, calcium phosphate co-precipitation and viral transduction for introducing DNA into cells are known in the art; therefore, the choice of method may lie with the competence and preference of the skilled practitioner. Examples of cells into which the IL4R α isogene may be introduced include, but are not limited to, continuous culture cells, such as COS, NIH/3T3, and primary or culture cells of the relevant tissue type, i.e., they express the IL4R α isogene. Such recombinant cells can be used to compare the biological activities of the different protein variants.

Recombinant organisms, i.e., transgenic animals, expressing a variant IL4R α gene are prepared using standard procedures known in the art. Preferably, a construct comprising the variant gene is introduced into a nonhuman animal or an ancestor of the animal at an embryonic stage, i.e., the one-cell stage, or generally not later than about the eight-cell stage. Transgenic animals carrying the constructs of the invention can be made by several methods known to those having skill in the art. One method involves transfecting into the embryo a retrovirus constructed to contain one or more insulator elements, a gene or genes of interest, and other components known to those skilled in the art to provide a complete shuttle vector harboring the insulated gene(s) as a transgene, see e.g., U.S. Patent No. 5,610,053. Another method involves directly injecting a transgene into the embryo. A third method involves the use

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of embryonic stem cells. Examples of animals into which the IL4Rα isogenes may be introduced include, but are not limited to, mice, rats, other rodents, and nonhuman primates (see "The Introduction of Foreign Genes into Mice" and the cited references therein, In: Recombinant DNA, Eds. J.D. Watson, M. Gilman, J. Witkowski, and M. Zoller; W.H. Freeman and Company, New York, pages 254-272). Transgenic animals stably expressing a human IL4Rα isogene and producing human IL4Rα protein can be used as biological models for studying diseases related to abnormal IL4Rα expression and/or activity, and for screening and assaying various candidate drugs, compounds, and treatment regimens to reduce the symptoms or effects of these diseases.

An additional embodiment of the invention relates to pharmaceutical compositions for treating disorders affected by expression or function of a novel IL4Ra isogene described herein. The pharmaceutical composition may comprise any of the following active ingredients: a polynucleotide comprising one of these novel IL4Ra isogenes; an antisense oligonucleotide directed against one of the novel IL4Ra isogenes, a polynucleotide encoding such an antisense oligonucleotide, or another compound which inhibits expression of a novel IL4R α isogene described herein. Preferably, the composition contains the active ingredient in a therapeutically effective amount. By therapeutically effective amount is meant that one or more of the symptoms relating to disorders affected by expression or function of a novel IL4Ra isogene is reduced and/or eliminated. The composition also comprises a pharmaceutically acceptable carrier, examples of which include, but are not limited to, saline, buffered saline, dextrose, and water. Those skilled in the art may employ a formulation most suitable for the active ingredient, whether it is a polynucleotide, oligonucleotide, protein, peptide or small molecule antagonist. The pharmaceutical composition may be administered alone or in combination with at least one other agent, such as a stabilizing compound. Administration of the pharmaceutical composition may be by any number of routes including, but not limited to oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, intradermal, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing Co., Easton, PA).

For any composition, determination of the therapeutically effective dose of active ingredient and/or the appropriate route of administration is well within the capability of those skilled in the art. For example, the dose can be estimated initially either in cell culture assays or in animal models. The animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans. The exact dosage will be determined by the practitioner, in light of factors relating to the patient requiring treatment, including but not limited to severity of the disease state, general health, age, weight and gender of the patient, diet, time and frequency of administration, other drugs being taken by the patient, and tolerance/response to the treatment.

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Information on the identity of genotypes and haplotypes for the IL4R α gene of any particular individual as well as the frequency of such genotypes and haplotypes in any particular population of individuals is expected to be useful for a variety of basic research and clinical applications. Thus, the invention also provides compositions and methods for detecting the novel IL4R α polymorphisms identified herein.

The compositions comprise at least one IL4Ra genotyping oligonucleotide. In one embodiment, an IL4R α genotyping oligonucleotide is a probe or primer capable of hybridizing to a target region that is located close to, or that contains, one of the novel polymorphic sites described herein. As used herein, the term "oligonucleotide" refers to a polynucleotide molecule having less than about 100 nucleotides. A preferred oligonucleotide of the invention is 10 to 35 nucleotides long. More preferably, the oligonucleotide is between 15 and 30, and most preferably, between 20 and 25 nucleotides in length. The oligonucleotide may be comprised of any phosphorylation state of ribonucleotides, deoxyribonucleotides, and acyclic nucleotide derivatives, and other functionally equivalent derivatives. Alternatively, oligonucleotides may have a phosphate-free backbone, which may be comprised of linkages such as carboxymethyl, acetamidate, carbamate, polyamide (peptide nucleic acid (PNA)) and the like (Varma, R. in Molecular Biology and Biotechnology, A Comprehensive Desk Reference, Ed. R. Meyers, VCH Publishers, Inc. (1995), pages 617-620). Oligonucleotides of the invention may be prepared by chemical synthesis using any suitable methodology known in the art, or may be derived from a biological sample, for example, by restriction digestion. The oligonucleotides may be labeled, according to any technique known in the art, including use of radiolabels, fluorescent labels, enzymatic labels, proteins, haptens, antibodies, sequence tags and the like.

Genotyping oligonucleotides of the invention must be capable of specifically hybridizing to a target region of an IL4R α polynucleotide, i.e., an IL4R α isogene. As used herein, specific hybridization means the oligonucleotide forms an anti-parallel double-stranded structure with the target region under certain hybridizing conditions, while failing to form such a structure when incubated with a non-target region or a non-IL4R α polynucleotide under the same hybridizing conditions. Preferably, the oligonucleotide specifically hybridizes to the target region under conventional high stringency conditions. The skilled artisan can readily design and test oligonucleotide probes and primers suitable for detecting polymorphisms in the IL4R α gene using the polymorphism information provided herein in conjunction with the known sequence information for the IL4R α gene and routine techniques.

A nucleic acid molecule such as an oligonucleotide or polynucleotide is said to be a "perfect" or "complete" complement of another nucleic acid molecule if every nucleotide of one of the molecules is complementary to the nucleotide at the corresponding position of the other molecule. A nucleic acid molecule is "substantially complementary" to another molecule if it hybridizes to that molecule with sufficient stability to remain in a duplex form under conventional low-stringency conditions.

Conventional hybridization conditions are described, for example, by Sambrook J. et al., in Molecular

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Cloning, A Laboratory Manual, 2nd Edition, Cold Spring Harbor Press, Cold Spring Harbor, NY (1989) and by Haymes, B.D. et al. in Nucleic Acid Hybridization, A Practical Approach, IRL Press, Washington, D.C. (1985). While perfectly complementary oligonucleotides are preferred for detecting polymorphisms, departures from complete complementarity are contemplated where such departures do not prevent the molecule from specifically hybridizing to the target region. For example, an oligonucleotide primer may have a non-complementary fragment at its 5' end, with the remainder of the primer being complementary to the target region. Alternatively, non-complementary nucleotides may be interspersed into the oligonucleotide probe or primer as long as the resulting probe or primer is still capable of specifically hybridizing to the target region.

Preferred genotyping oligonucleotides of the invention are allele-specific oligonucleotides. As used herein, the term allele-specific oligonucleotide (ASO) means an oligonucleotide that is able, under sufficiently stringent conditions, to hybridize specifically to one allele of a gene, or other locus, at a target region containing a polymorphic site while not hybridizing to the corresponding region in another allele(s). As understood by the skilled artisan, allele-specificity will depend upon a variety of readily optimized stringency conditions, including salt and formamide concentrations, as well as temperatures for both the hybridization and washing steps. Examples of hybridization and washing conditions typically used for ASO probes are found in Kogan et al., "Genetic Prediction of Hemophilia A" in PCR Protocols, A Guide to Methods and Applications, Academic Press, 1990 and Ruano et al., 87 Proc. Natl. Acad. Sci. USA 6296-6300, 1990. Typically, an allele-specific oligonucleotide will be perfectly complementary to one allele while containing a single mismatch for another allele.

Allele-specific oligonucleotide probes which usually provide good discrimination between different alleles are those in which a central position of the oligonucleotide probe aligns with the polymorphic site in the target region (e.g., approximately the 7^{th} or 8^{th} position in a 15 mer, the 8^{th} or 9^{th} position in a 16mer, the 10^{th} or 11^{th} position in a 20 mer). A preferred ASO probe for detecting IL4R α gene polymorphisms comprises a nucleotide sequence, listed 5' to 3', selected from the group consisting of:

```
(SEQ ID NO:4) and its complement,
          TTGCACCACTGCACT
                           (SEQ ID NO:5) and its complement,
          TTGCACCGCTGCACT
                           (SEQ ID NO:6) and its complement,
          TTTTGTGCTATTCCC
30
                           (SEQ ID NO:7) and its complement,
          TTTTGTGTTATTCCC
                           (SEQ ID NO:8) and its complement,
          CTGGGCCGCTCAGGC
                           (SEQ ID NO:9) and its complement,
          CTGGGCCTCTCAGGC
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          TAAGCCTGCGCTGGA (SEQ ID NO:10) and its complement,
          TAAGCCTCCGCTGGA (SEQ ID NO:11) and its complement,
          AGAACAACGGAGGCG (SEQ ID NO:12) and its complement,
          AGAACAATGGAGGCG (SEQ ID NO:13) and its complement,
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INDANSCE ALEGE
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(SEQ ID NO:52) and its complement,
         GCCTTCTTCCACCTT
                          (SEQ ID NO:53) and its complement,
         GCCTTCTCCCACCTT
                          (SEQ ID NO:54) and its complement,
         CAGCCCCGTCTCGG
                           (SEQ ID NO:55) and its complement,
5
         CAGCCCCTGTCTCGG
          GGAGTTTGTACATGC (SEQ ID NO:56) and its complement,
                          (SEQ ID NO:57) and its complement,
          GGAGTTTATACATGC
          CAGCTCCCCAGAGCA (SEQ ID NO:58) and its complement,
10
                          (SEQ ID NO:59) and its complement,
          CAGCTCCTCAGAGCA
                           (SEQ ID NO:60) and its complement,
          AGACAGGTCCTCGCC
                           (SEQ ID NO:61) and its complement,
          AGACAGGGCCTCGCC
15
          CTGCCCCTGGCAATG (SEQ ID NO:62) and its complement,
          CTGCCCCGGCAATG (SEQ ID NO:63) and its complement,
                           (SEQ ID NO:64) and its complement,
          AGGTGCATGTCCTCT
                           (SEQ ID NO:65) and its complement,
          AGGTGCACGTCCTCT
20
                          (SEQ ID NO:66) and its complement,
          GTGCATGTCCTCTTG
                          (SEQ ID NO:67) and its complement,
          GTGCATGCCCTCTTG
          GGCTTATCCATGCCT (SEQ ID NO:68) and its complement,
25
                           (SEQ ID NO:69) and its complement,
          GGCTTATTCATGCCT
                           (SEQ ID NO:70) and its complement,
          AGCCAGGCTGGCAGA
          AGCCAGGGTGGCAGA (SEQ ID NO:71) and its complement,
30
                           (SEQ ID NO:72) and its complement,
          GGCCCACATGGAGGC
                           (SEQ ID NO:73) and its complement,
          GGCCCACGTGGAGGC
          TAACACAGCCATCAA (SEQ ID NO:74) and its complement,
          TAACACAACCATCAA (SEQ ID NO:75) and its complement,
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          TAATGCTCGTCTGTG (SEQ ID NO:76) and its complement,
                           (SEQ ID NO:77) and its complement,
          TAATGCTTGTCTGTG
                           (SEQ ID NO:78) and its complement, and
          ACTTGCCGTCTGGGT
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```

ACTTGCCATCTGGGT

An allele-specific oligonucleotide primer of the invention has a 3' terminal nucleotide, or preferably a 3' penultimate nucleotide, that is complementary to only one nucleotide of a particular SNP, thereby acting as a primer for polymerase-mediated extension only if the allele containing that nucleotide is present. Allele-specific oligonucleotide primers hybridizing to either the coding or noncoding strand are contemplated by the invention. A preferred ASO primer for detecting IL4R α gene polymorphisms comprises a nucleotide sequence, listed 5' to 3', selected from the group consisting of:

(SEQ ID NO:79) and its complement.

```
CTGAGATTGCACCAC (SEQ ID NO:80); GGCTGGAGTGCAGTG (SEQ ID NO:81);

50 CTGAGATTGCACCGC (SEQ ID NO:82); GGCTGGAGTGCAGCG (SEQ ID NO:83);

CTGTGCTTTTGTGCT (SEQ ID NO:84); ACCAAGGGGAATAGC (SEQ ID NO:85);

CTGTGCTTTTGTGTT (SEQ ID NO:86); ACCAAGGGGAATAAC (SEQ ID NO:87);
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GAGTTCCTGGGCCGC (SEQ ID NO:88); GGAGCAGCCTGAGCG (SEQ ID NO:89);
    GAGTTCCTGGGCCTC (SEQ ID NO:90); GGAGCAGCCTGAGAG (SEQ ID NO:91);
    TCCGAGTAAGCCTGC (SEQ ID NO:92); TCCAGCTCCAGCGCA (SEQ ID NO:93);
    TCCGAGTAAGCCTCC (SEQ ID NO:94); TCCAGCTCCAGCGGA (SEQ ID NO:95);
    TCCCTGAGAACAACG (SEQ ID NO:96); ACCCCGCGCCTCCGT (SEQ ID NO:97);
    TCCCTGAGAACAATG (SEQ ID NO:98); ACCCCGCGCCTCCAT (SEQ ID NO:99);
    GAACAACGGAGGCGC (SEQ ID NO:100); CACACGCACCCCGCG (SEQ ID NO:101);
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    GAACAACGGAGGCAC (SEQ ID NO:102); CACACGCACCCCGTG (SEQ ID NO:103);
    TGGTCAGTGCGGATA (SEQ ID NO:104); CCAGTGTATAGTTAT (SEQ ID NO:105);
    TGGTCAGTGCGGACA (SEQ ID NO:106); CCAGTGTATAGTTGT (SEQ ID NO:107);
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    GCAGGGCGGAGTGCG (SEQ ID NO:108); AGCCACCCCTGCCGC (SEQ ID NO:109);
    GCAGGGCGGAGTGTG (SEQ ID NO:110); AGCCACCCCTGCCAC (SEQ ID NO:111);
    ACAGCTGCCTGGGCT (SEQ ID NO:112); CACCCCACCCTCAGC (SEQ ID NO:113);
    ACAGCTGCCTGGGTT (SEQ ID NO:114); CACCCCACCCTCAAC (SEQ ID NO:115);
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    TGAGGGTGGGGTGGG (SEQ ID NO:116); CCTCCTCCCCTGCCC (SEQ ID NO:117);
    TGAGGGTGGGGTGAG (SEQ ID NO:118); CCTCCTCCCCTGCTC (SEQ ID NO:119);
    GGCCGCTTCTCCCGC (SEQ ID NO:120); CTGGGTTTCACTGCG (SEQ ID NO:121);
    GGCCGCTTCTCCCAC (SEQ ID NO:122); CTGGGTTTCACTGTG (SEQ ID NO:123);
    TTTGGAGTGAAAACG (SEQ ID NO:124); CATCTGCCGGGTCGT (SEQ ID NO:125);
    TTTGGAGTGAAAATG (SEQ ID NO:126); CATCTGCCGGGTCAT (SEQ ID NO:127);
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    CTGGGAGGCAAGCCC (SEQ ID NO:128); TATCCAGCCCCAGGG (SEQ ID NO:129);
    CTGGGAGGCAAGCTC (SEQ ID NO:130); TATCCAGCCCCAGAG (SEQ ID NO:131);
    AGGCAAGCCCTGGGG (SEQ ID NO:132); TTTGCTATCCAGCCC (SEQ ID NO:133);
    AGGCAAGCCCTGGAG (SEQ ID NO:134); TTTGCTATCCAGCTC (SEQ ID NO:135);
35
    GGCTGGATAGCAAAT (SEQ ID NO:136); CTAGCTCCTGGGATT (SEQ ID NO:137);
     GGCTGGATAGCAATT (SEQ ID NO:138); CTAGCTCCTGGGAAT (SEQ ID NO:139);
     CACCTGGCTCTGCCC (SEQ ID NO:140); GGGACTTGCCTAGGG (SEQ ID NO:141);
40
     CACCTGGCTCTGCAC (SEQ ID NO:142); GGGACTTGCCTAGTG (SEQ ID NO:143);
     CCTGGCCCCCCACCC (SEQ ID NO:144); CTCTGATGTGAGGGG (SEQ ID NO:145);
     CCTGGCCCCCCACTC (SEQ ID NO:146); CTCTGATGTGAGGAG (SEQ ID NO:147);
45
     GAACCCTCCCTCCGC (SEQ ID NO:148); GCTGGCTGCGATGCG (SEQ ID NO:149);
     GAACCCTCCCTCCAC (SEQ ID NO:150); GCTGGCTGCGATGTG (SEQ ID NO:151);
     TAGATACACCTGCTG (SEQ ID NO:152); GCAGATACACCACAG (SEQ ID NO:153);
     TAGATACACCTGCCG (SEQ ID NO:154); GCAGATACACCACGG (SEQ ID NO:155);
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     GAAGGCATGTCTGAA (SEQ ID NO:156); ATGGCTGTCTACTTC (SEQ ID NO:157);
     GAAGGCATGTCTGCA (SEQ ID NO:158); ATGGCTGTCTACTGC (SEQ ID NO:159);
     GAACCCTGACCAACC (SEQ ID NO:160); TGCAAAAGCAAAGGA (SEQ ID NO:161);
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     GAACCCTGACCAATC (SEQ ID NO:162); TGCAAAAGCAAAGAA (SEQ ID NO:163);
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TCTTGCCCTGTTTTC (SEQ ID NO:164); TGTTGTGCTCCAGAA (SEQ ID NO:165);
    TCTTGCCCTGTTTCC (SEQ ID NO:166); TGTTGTGCTCCAGGA (SEQ ID NO:167);
    TGTTCCTGGACCTGC (SEQ ID NO:168); TCTCCTCTCCGAGCA (SEQ ID NO:169);
    TGTTCCTGGACCTTC (SEQ ID NO:170); TCTCCTCTCCGAGAA (SEQ ID NO:171);
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    TGGGGGAGTCATGCC (SEQ ID NO:172); AAGGTGGAAGAAGGC (SEQ ID NO:173);
    TGGGGGAGTCATGTC (SEQ ID NO:174); AAGGTGGAAGAAGAC (SEQ ID NO:175);
    AGTCATGCCTTCTTC (SEQ ID NO:176); TTCCCGAAGGTGGAA (SEQ ID NO:177);
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    AGTCATGCCTTCTCC (SEQ ID NO:178); TTCCCGAAGGTGGGA (SEQ ID NO:179);
    CAGCTGCAGCCCCCG (SEQ ID NO:180); TGGGGGCCGAGACGG (SEQ ID NO:181);
    CAGCTGCAGCCCCTG (SEQ ID NO:182); TGGGGGCCGAGACAG (SEQ ID NO:183);
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    CTATCAGGAGTTTGT (SEQ ID NO:184); TCCACCGCATGTACA (SEQ ID NO:185);
    CTATCAGGAGTTTAT (SEQ ID NO:186); TCCACCGCATGTATA (SEQ ID NO:187);
    CCCAAGCAGCTCCCC (SEQ ID NO:188); CCCAGGTGCTCTGGG (SEQ ID NO:189);
    CCCAAGCAGCTCCTC (SEQ ID NO:190); CCCAGGTGCTCTGAG (SEQ ID NO:191);
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    CTGTGGAGACAGGTC (SEQ ID NO:192); GTAGGGGGGGGAGGAC (SEQ ID NO:193);
    CTGTGGAGACAGGGC (SEQ ID NO:194); GTAGGGGGGGGGGCC (SEQ ID NO:195);
    TCCATCCTGCCCCTG (SEQ ID NO:196); TCTGAGCATTGCCAG (SEQ ID NO:197);
25
    TCCATCCTGCCCCG (SEQ ID NO:198); TCTGAGCATTGCCGG (SEQ ID NO:199);
    TCTCTTAGGTGCATG (SEQ ID NO:200); GCAACAAGAGGACAT (SEQ ID NO:201);
    TCTCTTAGGTGCACG (SEQ ID NO:202); GCAACAAGAGGACGT (SEQ ID NO:203);
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    TCTTAGGTGCATGTC (SEQ ID NO:204); CAGCAACAAGAGGAC (SEQ ID NO:205);
    TCTTAGGTGCATGCC (SEQ ID NO:206); CAGCAACAAGAGGGC (SEQ ID NO:207);
    GACTAGGGCTTATCC (SEQ ID NO:208); TTTCCCAGGCATGGA (SEQ ID NO:209);
    GACTAGGGCTTATTC (SEQ ID NO:210); TTTCCCAGGCATGAA (SEQ ID NO:211);
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    GAAGGCAGCCAGGCT (SEQ ID NO:212); TGGAAATCTGCCAGC (SEQ ID NO:213);
    GAAGGCAGCCAGGGT (SEQ ID NO:214); TGGAAATCTGCCACC (SEQ ID NO:215);
    GATCATGGCCCACAT (SEQ ID NO:216); AGGTGGGCCTCCATG (SEQ ID NO:217);
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     GATCATGGCCCACGT (SEQ ID NO:218); AGGTGGGCCTCCACG (SEQ ID NO:219);
     AGAAACTAACACAGC (SEQ ID NO:220); ATTCCCTTGATGGCT (SEQ ID NO:221);
     AGAAACTAACACAAC (SEQ ID NO:222); ATTCCCTTGATGGTT (SEQ ID NO:223);
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     GTTGAGTAATGCTCG (SEQ ID NO:224); AAAACACACAGACGA (SEQ ID NO:225);
     GTTGAGTAATGCTTG (SEQ ID NO:226); AAAACACACAGACAA (SEQ ID NO:227);
     TAAGAAACTTGCCGT (SEQ ID NO:228); ACCCAAACCCAGACG (SEQ ID NO:229);
     TAAGAAACTTGCCAT (SEQ ID NO:230); and ACCCAAACCCAGATG (SEQ ID NO:231).
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Other genotyping oligonucleotides of the invention hybridize to a target region located one to several nucleotides downstream of one of the novel polymorphic sites identified herein. Such oligonucleotides are useful in polymerase-mediated primer extension methods for detecting one of the

novel polymorphisms described herein and therefore such genotyping oligonucleotides are referred to herein as "primer-extension oligonucleotides". In a preferred embodiment, the 3′-terminus of a primer-extension oligonucleotide is a deoxynucleotide complementary to the nucleotide located immediately adjacent to the polymorphic site. A particularly preferred oligonucleotide primer for detecting IL4R α gene polymorphisms by primer extension terminates in a nucleotide sequence, listed 5′ to 3′, selected from the group consisting of:

•	AGATTGCACC	(SEQ ID NO:232);	TGGAGTGCAG	(SEQ ID NO:233);
	TGCTTTTGTG	(SEQ ID NO:234);	AAGGGGAATA	(SEQ ID NO:235);
	TTCCTGGGCC	(SEQ ID NO:236);	GCAGCCTGAG	(SEQ ID NO:237);
10	GAGTAAGCCT	(SEQ ID NO:238);	AGCTCCAGCG	(SEQ ID NO:239);
	CTGAGAACAA	(SEQ ID NO:240);	CCGCGCCTCC	(SEQ ID NO:241);
	CAACGGAGGC	(SEQ ID NO:242);	ACGCACCCCG	(SEQ ID NO:243);
	TCAGTGCGGA	(SEQ ID NO:244);	GTGTATAGTT	(SEQ ID NO:245);
	GGGCGGAGTG	(SEQ ID NO:246);	CACCCTGCC	(SEQ ID NO:247);
15	GCTGCCTGGG	(SEQ ID NO:248);	CCCACCCTCA	(SEQ ID NO:249);
	GGGTGGGGTG	(SEQ ID NO:250);	CCTCCCCTGC	(SEQ ID NO:251);
	CGCTTCTCCC	(SEQ ID NO:252);	GGTTTCACTG	(SEQ ID NO:253);
	GGAGTGAAAA	(SEQ ID NO:254);	CTGCCGGGTC	(SEQ ID NO:255);
	GGAGGCAAGC	(SEQ ID NO:256);	CCAGCCCCAG	(SEQ ID NO:257);
20	CAAGCCCTGG	(SEQ ID NO:258);	GCTATCCAGC	(SEQ ID NO:259);
	TGGATAGCAA	(SEQ ID NO:260);	GCTCCTGGGA	(SEQ ID-NO:261);
	CTGGCTCTGC	(SEQ ID NO:262);	ACTTGCCTAG	(SEQ ID NO:263);
*	GGCCCCCCAC	(SEQ ID NO:264);	TGATGTGAGG	(SEQ ID NO:265);
	CCCTCCCTCC	(SEQ ID NO:266);	GGCTGCGATG	(SEQ ID NO:267);
25	ATACACCTGC	(SEQ ID NO:268);	GATACACCAC	(SEQ ID NO:269);
	GGCATGTCTG	(SEQ ID NO:270);	GCTGTCTACT	(SEQ ID NO:271);
	CCCTGACCAA	(SEQ ID NO:272);	AAAAGCAAAG	(SEQ ID NO:273);
	TGCCCTGTTT	(SEQ ID NO:274);	TGTGCTCCAG	(SEQ ID NO:275);
	TCCTGGACCT	(SEQ ID NO:276);	CCTCTCCGAG	(SEQ ID NO:277);
30	GGGAGTCATG	(SEQ ID NO:278);	GTGGAAGAAG	(SEQ ID NO:279);
	CATGCCTTCT	(SEQ ID NO:280);	CCGAAGGTGG	(SEQ ID NO:281);
•	CTGCAGCCCC	(SEQ ID NO:282);	GGGCCGAGAC	(SEQ ID NO:283);
	TCAGGAGTTT	(SEQ ID NO:284);	ACCGCATGTA	(SEQ ID NO:285);
	AAGCAGCTCC	(SEQ ID NO:286);	AGGTGCTCTG	(SEQ ID NO:287);
35	TGGAGACAGG	(SEQ ID NO:288);	GGGGGCGAGG	(SEQ ID NO:289);
	ATCCTGCCCC	(SEQ ID NO:290);	GAGCATTGCC	(SEQ ID NO:291);
	CTTAGGTGCA	(SEQ ID NO:292);	ACAAGAGGAC	(SEQ ID NO:293);

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TAGGTGCATG	(SEQ ID	NO:294);	CAACAAGAGG	(SEQ	ID	NO:295);
TAGGGCTTAT	(SEQ ID	NO:296);	CCCAGGCATG	(SEQ	ID	NO:297);
GGCAGCCAGG	(SEQ ID	NO:298);	AAATCTGCCA	(SEQ	ID	NO:299);
CATGGCCCAC	(SEQ ID	NO:300);	TGGGCCTCCA	(SEQ	ID	NO:301);
AACTAACACA	(SEQ ID	NO:302);	CCCTTGATGG	(SEQ	ID	NO:303);
GAGTAATGCT	(SEQ ID	NO:304);	ACACACAGAC	(SEQ	ID	NO:305);
GAAACTTGCC	(SEQ ID	NO:306);	and CAAACCCAGA	(SEQ	ID	NO:307).

In some embodiments, a composition contains two or more differently labeled genotyping oligonucleotides for simultaneously probing the identity of nucleotides at two or more polymorphic sites. It is also contemplated that primer compositions may contain two or more sets of allele-specific primer pairs to allow simultaneous targeting and amplification of two or more regions containing a polymorphic site.

IL4R α genotyping oligonucleotides of the invention may also be immobilized on or synthesized on a solid surface such as a microchip, bead, or glass slide (see, e.g., WO 98/20020 and WO 98/20019). Such immobilized genotyping oligonucleotides may be used in a variety of polymorphism detection assays, including but not limited to probe hybridization and polymerase extension assays. Immobilized IL4R α genotyping oligonucleotides of the invention may comprise an ordered array of oligonucleotides designed to rapidly screen a DNA sample for polymorphisms in multiple genes at the same time.

In another embodiment, the invention provides a kit comprising at least two genotyping oligonucleotides packaged in separate containers. The kit may also contain other components such as hybridization buffer (where the oligonucleotides are to be used as a probe) packaged in a separate container. Alternatively, where the oligonucleotides are to be used to amplify a target region, the kit may contain, packaged in separate containers, a polymerase and a reaction buffer optimized for primer extension mediated by the polymerase, such as PCR.

The above described oligonucleotide compositions and kits are useful in methods for genotyping and/or haplotyping the IL4R α gene in an individual. As used herein, the terms "IL4R α genotype" and "IL4R α haplotype" mean the genotype or haplotype contains the nucleotide pair or nucleotide, respectively, that is present at one or more of the novel polymorphic sites described herein and may optionally also include the nucleotide pair or nucleotide present at one or more additional polymorphic sites in the IL4R α gene. The additional polymorphic sites may be currently known polymorphic sites or sites that are subsequently discovered.

One embodiment of the genotyping method involves isolating from the individual a nucleic acid mixture comprising the two copies of the IL4R α gene, or a fragment thereof, that are present in the individual, and determining the identity of the nucleotide pair at one or more of the polymorphic sites selected from PS1, PS2, PS3, PS4, PS6, PS7, PS8, PS9, PS10, PS11, PS12, PS13, PS14, PS15, PS16,

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PS17, PS18, PS19, PS20, PS21, PS22, PS23, PS25, PS27, PS28, PS30, PS32, PS33, PS34, PS35, PS36, PS37, PS38, PS39, PS40, PS41, PS44, and PS45 in the two copies to assign an ILARα genotype to the individual. As will be readily understood by the skilled artisan, the two "copies" of a gene in an individual may be the same allele or may be different alleles. In a preferred embodiment of the genotyping method, the identity of the nucleotide pair atone or more of the polymorphic sites selected from the group consisting of PS5, PS24, PS26, PS29, PS31, PS42, and PS43 is also determined. In a particularly preferred embodiment, the genotyping method comprises determining the identity of the nucleotide pair at each of PS1-45.

Typically, the nucleic acid mixture is isolated from a biological sample taken from the individual, such as a blood sample or tissue sample. Suitable tissue samples include whole blood, semen saliva, tears, urine, fecal material, sweat, buccal, skin and hair. The nucleic acid mixture may be comprised of genomic DNA, mRNA, or cDNA and, in the latter two cases, the biological sample must be obtained from an organ in which the IL4Rα gene is expressed. Furthermore it will be understood by the skilled artisan that mRNA or cDNA preparations would not be used to detect polymorphisms located in introns or in 5′ and 3′ nontranscribed regions. If an IL4Rα gene fragment is isolated, it must contain the polymorphic site(s) to be genotyped.

One embodiment of the haplotyping method comprises isolating from the individual a nucleic acid molecule containing only one of the two copies of the IL4Ra gene, or a fragment thereof, that is present in the individual and determining in that copy the identity of the nucleotide at one or more of the polymorphic sites PS1, PS2, PS3, PS4, PS6, PS7, PS8, PS9, PS10, PS11, PS12, PS13, PS14, PS15, PS16, PS17, PS18, PS19, PS20, PS21, PS22, PS23, PS25, PS27, PS28, PS30, PS32, PS33, PS34, PS35, PS36, PS37, PS38, PS39, PS40, PS41, PS44, and PS45 in that copy to assign an IL4Rα haplotype to the individual. The nucleic acid may be isolated using any method capable of separating the two copies of the IL4Ra gene or fragment such as one of the methods described above for preparing IL4Ra isogenes, with targeted in vivo cloning being the preferred approach. As will be readily appreciated by those skilled in the art, any individual clone will only provide haplotype information on one of the two IL4R α gene copies present in an individual. If haplotype information is desired for the individual's other copy, additional IL4Ra clones will need to be examined. Typically, at least five clones should be examined to have more than a 90% probability of haplotyping both copies of the IL4Ra gene in an individual. In some embodiments, the haplotyping method also comprises identifying the nucleotide atone or more of the polymorphic sites PS5, PS24, PS26, PS29, PS31, PS42, and PS43. In a particularly preferred embodiment, the nucleotide at each of PS1-45 is identified.

In a preferred embodiment, an IL4R α haplotype pair is determined for an individual by identifying the phased sequence of nucleotides at one or more of the polymorphic sites selected from PS1, PS2, PS3, PS4, PS6, PS7, PS8, PS9, PS10, PS11, PS12, PS13, PS14, PS15, PS16, PS17, PS18, PS19, PS20, PS21, PS22, PS23, PS25, PS27, PS28, PS30, PS32, PS33, PS34, PS35, PS36, PS37, PS38,

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PS39, PS40, PS41, PS44, and PS45 in each copy of the IL4Rα gene that is present in the individual. In a particularly preferred embodiment, the haplotyping method comprises identifying the phased sequence of nucleotides at each of PS1-45 in each copy of the IL4Rα gene. When haplotyping both copies of the gene, the identifying step is preferably performed with each copy of the gene being placed in separate containers. However, it is also envisioned that if the two copies are labeled with different tags, or are otherwise separately distinguishable or identifiable, it could be possible in some cases to perform the method in the same container. For example, if first and second copies of the gene are labeled with different first and second fluorescent dyes, respectively, and an allele-specific oligonucleotide labeled with yet a third different fluorescent dye is used to assay the polymorphic site(s), then detecting a combination of the first and third dyes would identify the polymorphism in the first gene copy while detecting a combination of the second and third dyes would identify the polymorphism in the second gene copy.

In both the genotyping and haplotyping methods, the identity of a nucleotide (or nucleotide pair) at a polymorphic site(s) may be determined by amplifying a target region(s) containing the polymorphic site(s) directly from one or both copies of the IL4R α gene, or fragment thereof, and the sequence of the amplified region(s) determined by conventional methods. It will be readily appreciated by the skilled artisan that only one nucleotide will be detected at a polymorphic site in individuals who are homozygous at that site, while two different nucleotides will be detected if the individual is heterozygous for that site. The polymorphism may be identified directly, known as positive-type identification, or by inference, referred to as negative-type identification. For example, where a SNP is known to be guanine and cytosine in a reference population, a site may be positively determined to be either guanine or cytosine for an individual homozygous at that site, or both guanine and cytosine, if the individual is heterozygous at that site. Alternatively, the site may be negatively determined to be not guanine (and thus cytosine/cytosine) or not cytosine (and thus guanine/guanine).

In addition, the identity of the allele(s) present at any of the novel polymorphic sites described herein may be indirectly determined by genotyping a polymorphic site not disclosed herein that is in linkage disequilibrium with the polymorphic site that is of interest. Two sites are said to be in linkage disequilibrium if the presence of a particular variant at one site enhances the predictability of another variant at the second site (Stevens, JC 1999, *Mol. Diag.* 4: 309-17). Polymorphic sites in linkage disequilibrium with the presently disclosed polymorphic sites may be located in regions of the gene or in other genomic regions not examined herein. Genotyping of a polymorphic site in linkage disequilibrium with the novel polymorphic sites described herein may be performed by, but is not limited to, any of the above-mentioned methods for detecting the identity of the allele at a polymorphic site.

The target region(s) may be amplified using any oligonucleotide-directed amplification method, including but not limited to polymerase chain reaction (PCR) (U.S. Patent No. 4,965,188), ligase chain reaction (LCR) (Barany et al., Proc. Natl. Acad. Sci. USA 88:189-193, 1991; WO90/01069), and oligonucleotide ligation assay (OLA) (Landegren et al., Science 241:1077-1080, 1988).

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Oligonucleotides useful as primers or probes in such methods should specifically hybridize to a region of the nucleic acid that contains or is adjacent to the polymorphic site. Typically, the oligonucleotides are between 10 and 35 nucleotides in length and preferably, between 15 and 30 nucleotides in length. Most preferably, the oligonucleotides are 20 to 25 nucleotides long. The exact length of the oligonucleotide will depend on many factors that are routinely considered and practiced by the skilled artisan.

Other known nucleic acid amplification procedures may be used to amplify the target region including transcription-based amplification systems (U.S. Patent No. 5,130,238; EP 329,822; U.S. Patent No. 5,169,766, WO89/06700) and isothermal methods (Walker et al., *Proc. Natl. Acad. Sci. USA* 89:392-396, 1992).

A polymorphism in the target region may also be assayed before or after amplification using one of several hybridization-based methods known in the art. Typically, allele-specific oligonucleotides are utilized in performing such methods. The allele-specific oligonucleotides may be used as differently labeled probe pairs, with one member of the pair showing a perfect match to one variant of a target sequence and the other member showing a perfect match to a different variant. In some embodiments, more than one polymorphic site may be detected at once using a set of allele-specific oligonucleotides or oligonucleotide pairs. Preferably, the members of the set have melting temperatures within 5°C, and more preferably within 2°C, of each other when hybridizing to each of the polymorphic sites being detected.

Hybridization of an allele-specific oligonucleotide to a target polynucleotide may be performed with both entities in solution, or such hybridization may be performed when either the oligonucleotide or the target polynucleotide is covalently or noncovalently affixed to a solid support. Attachment may be mediated, for example, by antibody-antigen interactions, poly-L-Lys, streptavidin or avidin-biotin, salt bridges, hydrophobic interactions, chemical linkages, UV cross-linking baking, etc. Allele-specific oligonucleotides may be synthesized directly on the solid support or attached to the solid support subsequent to synthesis. Solid-supports suitable for use in detection methods of the invention include substrates made of silicon, glass, plastic, paper and the like, which may be formed, for example, into wells (as in 96-well plates), slides, sheets, membranes, fibers, chips, dishes, and beads. The solid support may be treated, coated or derivatized to facilitate the immobilization of the allele-specific oligonucleotide or target nucleic acid.

The genotype or haplotype for the IL4R α gene of an individual may also be determined by hybridization of a nucleic sample containing one or both copies of the gene to nucleic acid arrays and subarrays such as described in WO 95/11995. The arrays would contain a battery of allele-specific oligonucleotides representing each of the polymorphic sites to be included in the genotype or haplotype.

The identity of polymorphisms may also be determined using a mismatch detection technique, including but not limited to the RNase protection method using riboprobes (Winter et al., Proc. Natl. Acad. Sci. USA 82:7575, 1985; Meyers et al., Science 230:1242, 1985) and proteins which recognize nucleotide mismatches, such as the E. coli mutS protein (Modrich, P. Ann. Rev. Genet. 25:229-253,

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1991). Alternatively, variant alleles can be identified by single strand conformation polymorphism (SSCP) analysis (Orita et al., Genomics 5:874-879, 1989; Humphries et al., in Molecular Diagnosis of Genetic Diseases, R. Elles, ed., pp. 321-340, 1996) or denaturing gradient gel electrophoresis (DGGE) (Wartell et al., Nucl. Acids Res. 18:2699-2706, 1990; Sheffield et al., Proc. Natl. Acad. Sci. USA 86:232-236, 1989).

A polymerase-mediated primer extension method may also be used to identify the polymorphism(s). Several such methods have been described in the patent and scientific literature and include the "Genetic Bit Analysis" method (WO92/15712) and the ligase/polymerase mediated genetic bit analysis (U.S. Patent 5,679,524. Related methods are disclosed in WO91/02087, WO90/09455, WO95/17676, U.S. Patent Nos. 5,302,509, and 5,945,283. Extended primers containing a polymorphism may be detected by mass spectrometry as described in U.S. Patent No. 5,605,798. Another primer extension method is allele-specific PCR (Ruaño et al., Nucl. Acids Res. 17:8392, 1989; Ruaño et al., Nucl. Acids Res. 19, 6877-6882, 1991; WO 93/22456; Turki et al., J. Clin. Invest. 95:1635-1641, 1995). In addition, multiple polymorphic sites may be investigated by simultaneously amplifying multiple regions of the nucleic acid using sets of allele-specific primers as described in Wallace et al. (WO89/10414).

In another aspect of the invention, an individual's IL4Rα haplotype pair is predicted from its IL4Rα genotype using information on haplotype pairs known to exist in a reference population. In its broadest embodiment, the haplotyping prediction method comprises identifying an IL4Rα genotype for the individual at two or more polymorphic sites selected from PS1, PS2, PS3, PS4, PS6, PS7, PS8, PS9, PS10, PS11, PS12, PS13, PS14, PS15, PS16, PS17, PS18, PS19, PS20, PS21, PS22, PS23, PS25, PS27, PS28, PS30, PS32, PS33, PS34, PS35, PS36, PS37, PS38, PS39, PS40, PS41, PS44, and PS45, enumerating all possible haplotype pairs which are consistent with the genotype, accessing data containing IL4Rα haplotype pairs identified in a reference population, and assigning a haplotype pair to the individual that is consistent with the data. In one embodiment, the reference haplotype pairs include the IL4Rα haplotype pairs shown in Table 4.

Generally, the reference population should be composed of randomly-selected individuals representing the major ethnogeographic groups of the world. A preferred reference population for use in the methods of the present invention comprises an approximately equal number of individuals from Caucasian, African American, Asian and Hispanic-Latino population groups with the minimum number of each group being chosen based on how rare a haplotype one wants to be guaranteed to see. For example, if one wants to have a q% chance of not missing a haplotype that exists in the population at a p% frequency of occurring in the reference population, the number of individuals (n) who must be sampled is given by $2n=\log(1-q)/\log(1-p)$ where p and q are expressed as fractions. A preferred reference population allows the detection of any haplotype whose frequency is at least 10% with about 99% certainty and comprises about 20 unrelated individuals from each of the four population groups named above. A particularly preferred reference population includes a 3-generation family representing

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one or more of the four population groups to serve as controls for checking quality of haplotyping procedures.

In a preferred embodiment, the haplotype frequency data for each ethnogeographic group is examined to determine whether it is consistent with Hardy-Weinberg equilibrium. Hardy-Weinberg equilibrium (D.L. Hartl et al., Principles of Population Genomics, Sinauer Associates (Sunderland, MA), 3^{rd} Ed., 1997) postulates that the frequency of finding the haplotype pair H_1/H_2 is equal to $p_{H-W}(H_1/H_2) = 2p(H_1)p(H_2)$ if $H_1 \neq H_2$ and $p_{H-W}(H_1/H_2) = p(H_1)p(H_2)$ if $H_1 = H_2$. A statistically significant difference between the observed and expected haplotype frequencies could be due to one or more factors including significant inbreeding in the population group, strong selective pressure on the gene, sampling bias, and/or errors in the genotyping process. If large deviations from Hardy-Weinberg equilibrium are observed in an ethnogeographic group, the number of individuals in that group can be increased to see if the deviation is due to a sampling bias. If a larger sample size does not reduce the difference between observed and expected haplotype pair frequencies, then one may wish to consider haplotyping the individual using a direct haplotyping method such as, for example, CLASPER System technology (U.S. Patent No. 5,866,404), SMD, or allele-specific long-range PCR (Michalotos-Beloin et al., Nucleic Acids Res. 24:4841-4843, 1996).

In one embodiment of this method for predicting an IL4R α haplotype pair, the assigning step involves performing the following analysis. First, each of the possible haplotype pairs is compared to the haplotype pairs in the reference population. Generally, only one of the haplotype pairs in the reference population matches a possible haplotype pair and that pair is assigned to the individual. Occasionally, only one haplotype represented in the reference haplotype pairs is consistent with a possible haplotype pair for an individual, and in such cases the individual is assigned a haplotype pair containing this known haplotype and a new haplotype derived by subtracting the known haplotype from the possible haplotype pair. In rare cases, either no haplotypes in the reference population are consistent with the possible haplotype pairs, or alternatively, multiple reference haplotype pairs are consistent with the possible haplotype pairs. In such cases, the individual is preferably haplotyped using a direct molecular haplotyping method such as, for example, CLASPER System[™] technology (U.S. Patent No. 5,866,404), SMD, or allele-specific long-range PCR (Michalotos-Beloin et al., Nucleic Acids Res. 24:4841-4843, 1996).

The invention also provides a method for determining the frequency of an IL4R α genotype or IL4R α haplotype in a population. The method comprises determining the genotype or the haplotype pair for the IL4R α gene that is present in each member of the population, wherein the genotype or haplotype comprises the nucleotide pair or nucleotide detected at one or more of the polymorphic sites PS1, PS2, PS3, PS4, PS6, PS7, PS8, PS9, PS10, PS11, PS12, PS13, PS14, PS15, PS16, PS17, PS18, PS19, PS20, PS21, PS22, PS23, PS25, PS27, PS28, PS30, PS32, PS33, PS34, PS35, PS36, PS37, PS38, PS39, PS40, PS41, PS44, and PS45 in the IL4R α gene; and calculating the frequency any particular genotype or

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haplotype is found in the population. The population may be a reference population, a family population, a same sex population, a population group, a trait population (e.g., a group of individuals exhibiting a trait of interest such as a medical condition or response to a therapeutic treatment).

In another aspect of the invention, frequency data for IL4Ra genotypes and/or haplotypes found in a reference population are used in a method for identifying an association between a trait and an IL4R α genotype or an IL4R α haplotype. The trait may be any detectable phenotype, including but not limited to susceptibility to a disease or response to a treatment. The method involves obtaining data on the frequency of the genotype(s) or haplotype(s) of interest in a reference population as well as in a population exhibiting the trait. Frequency data for one or both of the reference and trait populations may be obtained by genotyping or haplotyping each individual in the populations using one of the methods described above. The haplotypes for the trait population may be determined directly or, alternatively, by the predictive genotype to haplotype approach described above. In another embodiment, the frequency data for the reference and/or trait populations is obtained by accessing previously determined frequency data, which may be in written or electronic form. For example, the frequency data may be present in a database that is accessible by a computer. Once the frequency data is obtained, the frequencies of the genotype(s) or haplotype(s) of interest in the reference and trait populations are compared. In a preferred embodiment, the frequencies of all genotypes and/or haplotypes observed in the populations are compared. If a particular genotype or haplotype for the IL4R α gene is more frequent in the trait population than in the reference population at a statistically significant amount, then the trait is predicted to be associated with that IL4R α genotype or haplotype. Preferably, the IL4R α genotype or haplotype being compared in the trait and reference populations is selected from the full-genotypes and fullhaplotypes shown in Tables 4 and 5, respectively, or from sub-genotypes and sub-haplotypes derived from these genotypes and haplotypes.

In a preferred embodiment of the method, the trait of interest is a clinical response exhibited by a patient to some therapeutic treatment, for example, response to a drug targeting IL4R α or response to a therapeutic treatment for a medical condition. As used herein, "medical condition" includes but is not limited to any condition or disease manifested as one or more physical and/or psychological symptoms for which treatment is desirable, and includes previously and newly identified diseases and other disorders. As used herein the term "clinical response" means any or all of the following: a quantitative measure of the response, no response, and adverse response (i.e., side effects).

In order to deduce a correlation between clinical response to a treatment and an IL4R α genotype or haplotype, it is necessary to obtain data on the clinical responses exhibited by a population of individuals who received the treatment, hereinafter the "clinical population". This clinical data may be obtained by analyzing the results of a clinical trial that has already been run and/or the clinical data may be obtained by designing and carrying out one or more new clinical trials. As used herein, the term "clinical trial" means any research study designed to collect clinical data on responses to a particular

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treatment, and includes but is not limited to phase I, phase II and phase III clinical trials. Standard methods are used to define the patient population and to enroll subjects.

It is preferred that the individuals included in the clinical population have been graded for the existence of the medical condition of interest. This is important in cases where the symptom(s) being presented by the patients can be caused by more than one underlying condition, and where treatment of the underlying conditions are not the same. An example of this would be where patients experience breathing difficulties that are due to either asthma or respiratory infections. If both sets were treated with an asthma medication, there would be a spurious group of apparent non-responders that did not actually have asthma. These people would affect the ability to detect any correlation between haplotype and treatment outcome. This grading of potential patients could employ a standard physical exam or one or more lab tests. Alternatively, grading of patients could use haplotyping for situations where there is a strong correlation between haplotype pair and disease susceptibility or severity.

The therapeutic treatment of interest is administered to each individual in the trial population and each individual's response to the treatment is measured using one or more predetermined criteria. It is contemplated that in many cases, the trial population will exhibit a range of responses and that the investigator will choose the number of responder groups (e.g., low, medium, high) made up by the various responses. In addition, the IL4R α gene for each individual in the trial population is genotyped and/or haplotyped, which may be done before or after administering the treatment.

After both the clinical and polymorphism data have been obtained, correlations between individual response and IL4R α genotype or haplotype content are created. Correlations may be produced in several ways. In one method, individuals are grouped by their IL4R α genotype or haplotype (or haplotype pair) (also referred to as a polymorphism group), and then the averages and standard deviations of clinical responses exhibited by the members of each polymorphism group are calculated.

These results are then analyzed to determine if any observed variation in clinical response between polymorphism groups is statistically significant. Statistical analysis methods which may be used are described in L.D. Fisher and G. vanBelle, "Biostatistics: A Methodology for the Health Sciences", Wiley-Interscience (New York) 1993. This analysis may also include a regression calculation of which polymorphic sites in the IL4R α gene give the most significant contribution to the differences in phenotype. One regression model useful in the invention starts with a model of the form

$$r = r_0 + S \times d$$

where r is the response, r_0 is a constant called the "intercept", S is the slope and d is the dose. To determine the dose, the most-common and least common nucleotides at the polymorphic site are first defined. Then, for each individual in the trial population, one calculates a "dose" as the number of least-common nucleotides the individual has at the polymorphic site of interest. This value can be 0 (homozygous for the least-common nucleotide), 1 (heterozygous), or 2 (homozygous for the most common nucleotide). An individual's "response" is the value of the clinical measurement. Standard

linear regression methods are then used to fit all the individuals' doses and responses to a single model (see e.g., L.D. Fisher and G. vanBelle, supra, Ch 9). The outputs of the regression calculation are the intercept r_0 , the slope S, and the variance (which measures how well the data fits this simple linear model). The Students t-test value and the level of significance can then be calculated for each of the polymorphic sites.

A second method for finding correlations between IL4Rα haplotype content and clinical responses uses predictive models based on error-minimizing optimization algorithms. One of many possible optimization algorithms is a genetic algorithm (R. Judson, "Genetic Algorithms and Their Uses in Chemistry" in Reviews in Computational Chemistry, Vol. 10, pp. 1-73, K. B. Lipkowitz and D. B. Boyd, eds. (VCH Publishers, New York, 1997). Simulated annealing (Press et al., "Numerical Recipes in C: The Art of Scientific Computing", Cambridge University Press (Cambridge) 1992, Ch. 10), neural networks (E. Rich and K. Knight, "Artificial Intelligence", 2nd Edition (McGraw-Hill, New York, 1991, Ch. 18), standard gradient descent methods (Press et al., supra Ch. 10), or other global or local optimization approaches (see discussion in Judson, supra) could also be used. As an example, a genetic algorithm approach is described herein. This method searches for optimal parameters or weights in linear or non-linear models connecting IL4Rα haplotype loci and clinical outcome. One model is of the form

$$C = C_0 + \sum_{\alpha} \left(\sum_{i} w_{i,\alpha} R_{i,\alpha} + \sum_{i} w'_{i,\alpha} L_{i,\alpha} \right)$$
 [1]

where C is the measured clinical outcome, i goes over all polymorphic sites, α over all candidate genes, C_0 ; $w_{i,\alpha}$ and $w'_{i,\alpha}$ are variable weight values, $R_{i,\alpha}$ is equal to 1 if site i in gene α in the first haplotype takes on the most common nucleotide and -1 if it takes on the less common nucleotide. $L_{i,\alpha}$ is the same as $R_{i,\alpha}$ except for the second haplotype. The constant term C_0 and the weights $w_{i,\alpha}$ and $w'_{i,\alpha}$ are varied by the genetic algorithm during a search process that minimizes the error between the measured value of C and the value calculated from Equation 1. Models other than the one given in Equation 1 can be readily incorporated by those skilled in the art for analyzing the clinical and polymorphism data. The genetic algorithm is especially suited for searching not only over the space of weights in a particular model but also over the space of possible models (Judson, supra).

Correlations may also be analyzed using analysis of variation (ANOVA) techniques to determine how much of the variation in the clinical data is explained by different subsets of the polymorphic sites in the IL4R α gene. ANOVA is used to test hypotheses about whether a response variable is caused by or correlated with one or more traits or variables that can be measured (Fisher and vanBelle, supra, Ch. 10). These traits or variables are called the independent variables. To carry out ANOVA, the independent variable(s) are measured and individuals are placed into groups based on their values for these variables. In this case, the independent variable(s) refers to the combination of polymorphisms present at a subset of the polymorphic sites, and thus, each group contains those individuals with a given genotype or

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haplotype pair. The variation in response within the groups and also the variation between groups is then measured. If the within-group response variation is large (people in a group have a wide range of responses) and the response variation between groups is small (the average responses for all groups are about the same) then it can be concluded that the independent variables used for the grouping are not causing or correlated with the response variable. For instance, if people are grouped by month of birth (which should have nothing to do with their response to a drug) the ANOVA calculation should show a low level of significance. However, if the response variation is larger between groups than within groups, the F-ratio (="between groups" divided by "within groups") is greater than one. Large values of the F-ratio indicate that the independent variable is causing or correlated with the response. The calculated F-ratio is preferably compared with the critical F-distribution value at whatever level of significance is of interest. If the F-ratio is greater than the Critical F-distribution value, then one may be confident that the individual's genotype or haplotype pair for this particular subset of polymorphic sites in the IL4Rα gene is at least partially responsible for, or is at least strongly correlated with the clinical response.

From the analyses described above, a mathematical model may be readily constructed by the skilled artisan that predicts clinical response as a function of IL4R α genotype or haplotype content. Preferably, the model is validated in one or more follow-up clinical trials designed to test the model.

The identification of an association between a clinical response and a genotype or haplotype (or haplotype pair) for the IL4R α gene may be the basis for designing a diagnostic method to determine those individuals who will or will not respond to the treatment, or alternatively, will respond at a lower level and thus may require more treatment, i.e., a greater dose of a drug. The diagnostic method may take one of several forms: for example, a direct DNA test (i.e., genotyping or haplotyping one or more of the polymorphic sites in the IL4R α gene), a serological test, or a physical exam measurement. The only requirement is that there be a good correlation between the diagnostic test results and the underlying IL4R α genotype or haplotype that is in turn correlated with the clinical response. In a preferred embodiment, this diagnostic method uses the predictive haplotyping method described above.

Any or all analytical and mathematical operations involved in practicing the methods of the present invention may be implemented by a computer. In addition, the computer may execute a program that generates views (or screens) displayed on a display device and with which the user can interact to view and analyze large amounts of information relating to the IL4R α gene and its genomic variation, including chromosome location, gene structure, and gene family, gene expression data, polymorphism data, genetic sequence data, and clinical data population data (e.g., data on ethnogeographic origin, clinical responses, genotypes, and haplotypes for one or more populations). The IL4R α polymorphism data described herein may be stored as part of a relational database (e.g., an instance of an Oracle database or a set of ASCII flat files). These polymorphism data may be stored on the computer's hard drive or may, for example, be stored on a CD ROM or on one or more other storage devices accessible

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by the computer. For example, the data may be stored on one or more databases in communication with the computer via a network.

Preferred embodiments of the invention are described in the following examples. Other embodiments within the scope of the claims herein will be apparent to one skilled in the art from consideration of the specification or practice of the invention as disclosed herein. It is intended that the specification, together with the examples, be considered exemplary only, with the scope and spirit of the invention being indicated by the claims which follow the examples.

EXAMPLES

The Examples herein are meant to exemplify the various aspects of carrying out the invention and are not intended to limit the scope of the invention in any way. The Examples do not include detailed descriptions for conventional methods employed, such as in the performance of genomic DNA isolation, PCR and sequencing procedures. Such methods are well-known to those skilled in the art and are described in numerous publications, for example, Sambrook, Fritsch, and Maniatis, "Molecular Cloning: A Laboratory Manual", 2nd Edition, Cold Spring Harbor Laboratory Press, USA, (1989).

Example 1A

This example illustrates examination of various regions of the IL4R α gene for polymorphic sites using DNA from Index Repository IA.

Amplification of Target Regions

The following target regions of the IL4R α gene were amplified using the PCR primer pairs listed below, with the sequences presented in the 5' to 3' direction and nucleotide positions shown for each region corresponding to the indicated GenBank Accession No.

25 Accession Number: AC004525

Fragment 1

Forward Primer

32801-32822 CCACAGTCATCCCGACACTAGC (SEQ ID NO:308)

Reverse Primer

30 Complement of 33355-33334 TATTCCAGCCGTATCCATGTGC (SEQ ID NO:309)

PCR product 555 nt

Fragment 2

Forward Primer

35 35515-35536 CCTTGGTGCATGTGGTAAGAGG (SEQ ID NO:310)

Reverse Primer

Complement of 36068-36046 TTTCAAAGGTGGGAGGACTGAGG (SEQ ID NO:311)

PCR product 554 nt

40 Fragment 3
Forward Primer

37031-37050 GCAGTGAGCTGGGATTGTGC (SEQ ID NO:312)

Reverse Primer	•
Complement of 37701-37679	AACTCCCCTTCTCTGATGTGAGG (SEQ ID NO:313)
PCR product 671 nt	

Fragment 4
Forward Primer
43240-43262 TCACAGTTACAGAGGTGGCAAGC (SEQ ID NO:314)
Reverse Primer
Complement of 43727-43706 CTGCCTACCTGGCAGATACACC (SEQ ID NO:315)

10 PCR product 488 nt

Fragment 5
Forward Primer
49553-49574 AGCTGTCACTCCACCTCCTTGG (SEQ ID NO:316)

15 Reverse Primer
Complement of 50036-50013 AAAGCCTCTGGTCTGCTAATGACC (SEQ ID NO:317)
PCR product 484 nt

Fragment 6
20 Forward Primer
51394-51415 GGGAGGAGATTCAGAGCACTCC (SEQ ID NO:318)
Reverse Primer
Complement of 51847-51826 CAGTCCACGTTTCCAGAACACC (SEQ ID NO:319)
PCR product 454 nt

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Fragment 7
Forward Primer
52806-52826 GGCTTGGGATAATGGTGTTGC (SEQ ID NO:320)
Reverse Primer

Complement of 53529-53507 TACTTCCCGAAGGTGGAAGAAGG (SEQ ID NO:321)
PCR product 724 nt

Fragment 8 Forward Primer

53242-53265 CAGTGGAGATCAGCAAGACAGTCC (SEQ ID NO:322)
Reverse Primer
Complement of 53807-53786 GGGCATCTCGGGTTCTACTTCC (SEQ ID NO:323)
PCR product 566 nt

40 Fragment 9
Forward Primer
53522-53544 GGGAAGTACGAGTGCTCACATGC (SEQ ID NO:324)
Reverse Primer
Complement of 54110-54088 CTTATACCCCTCTTCCCCACTGC (SEQ ID NO:325)
45 PCR product 589 nt

Fragment 10
Forward Primer
53821-53841 TCTCTGAGCCAACCACTGTGC (SEQ ID NO:326)

Reverse Primer
Complement of 54359-54337 GGCTGAGTAGACAATGCCACTGC (SEQ ID NO:327)
PCR product 539 nt

5	Fragment 11 Forward Primer 54055-54076 CTGTGTCCCCAGAGAAATGTGG (SEQ ID NO:328) Reverse Primer Complement of 54717-54695 GACTCAGCAACAAGAGGACATGC (SEQ ID NO:328) PCR product 663 nt	29)
10	Fragment 12 Forward Primer 54342-54365 GGCATTGTCTACTCAGCCCTTACC (SEQ ID NO:330) Reverse Primer Complement of 54986-54967 ACAAGTCGAGGTGCCCAAGG (SEQ ID NO:331) PCR product 645 nt	
15	Fragment 13 Forward Primer 54669-54693 CCCACATACATGAGGGTCTCTTAGG (SEQ ID NO:332) Reverse Primer Complement of 55270-55250 ATTCTGCCTCCAGCATCAACC (SEQ ID NO:333) PCR product 602 nt	
25	Fragment 14 Forward Primer 55235-55258 AACAGAGCTTCCTTAGGTTGATGC (SEQ ID NO:334) Reverse Primer Complement of 55847-55825 CCTCAGTTCCCCACTACCTTAGC (SEQ ID NO:335) PCR product 613 nt	5)
	These primer pairs were used in PCR reactions containing genomic DNA isolated	d from
30	immortalized cell lines for each member of Index Repository IA. The PCR reactions we	
	under the following conditions:	
35	Reaction volume 10 x Advantage 2 Polymerase reaction buffer (Clontech) 100 ng of human genomic DNA 10 mM dNTP Advantage 2 Polymerase enzyme mix (Clontech) Forward Primer (10 µM)	= 20 μ l = 2 μ l = 1 μ l = 0.4 μ l = 0.2 μ l = 0.4 μ l = 0.4 μ l
	Reverse Primer (10 μM) Water	= $0.4 \mu l$ = $15.6 \mu l$
40	Amplification profile: 94°C - 2 min. 1 cycle	
45	94°C - 30 sec. 70°C - 45 sec. 72°C - 1 min. } 10 cycles	
50	94°C - 30 sec. 64°C - 45 sec. 72°C - 1 min. 35 cycles	

Sequencing of PCR Products

The PCR products were purified by Solid Phase Reversible Immobilization using the protocol developed by the Whitehead Genome Center. A detailed protocol can be found at http://www.genome.wi.mit.edu/sequencing/protocols/pure/SPRI_pcr.html.

Briefly, five μl of carboxyl coated magnetic beads (10 mg/ml) and 60 μl of HYB BUFFER (2.5M NaCl/20% PEG 8000) were added to each PCR reaction mixture (20 μl). The reaction mixture was mixed well and incubated at room temperature (RT) for 10 min. The microtitre plate was placed on a magnet for 2 min and the beads washed twice with 150 μl of 70% EtOH. The beads were air dried for 2 min and the DNA was eluted in 25 μl of distilled water and incubated at RT for 5 min. The beads were magnetically separated and the supernatant removed for testing and sequencing.

The purified PCR products were sequenced in both directions using the primer sets described previously or those listed, in the 5' to 3' direction, below.

Accession Number: AC004525

Fragment 1

Forward Primer

32865-32882 GCGCTGGCCCTCAACTTT (SEQ ID NO:336)

Reverse Primer

Complement of 33283-33264 GTCCCTGGAGATGGGACCTC (SEQ ID NO:337)

Fragment 2

Forward Primer

35598-35617 GCCCCCAGATCTGTCCTCAC (SEQ ID NO:338)

Reverse Primer

Complement of 36013-35994 GGAAAATACAGGCGGCTTCC (SEQ ID NO:339)

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Fragment 3

Forward Primer

37182-37203 GGCTCTGAATCTGTGTGTGCT (SEQ ID NO:340)

Reverse Primer

30 Complement of 37639-37620 AGCCAGGTGAGAAGCCAGGT (SEQ ID NO:341)

Fragment 4

Forward Primer

43266-43285 GGCCTGAACAGGACGAACAA (SEQ ID NO:342)

35 Reverse Primer

Complement of 43687-43668 GGCAGGATTGCCATTAGAGG (SEQ ID NO:343)

Fragment 5

Forward Primer

40 49639-49660 TGAGTCAGTGGTTTGACCTCCA (SEQ ID NO:344)

Reverse Primer

Complement of 49999-49980 GCCTCTGTCTCCCCTGCAAC (SEQ ID NO:345)

Fragment 6

45 Forward Primer

51423-51442 CCACTTTTGCCATCGACCAC (SEQ ID NO:346)

Reverse Primer

Complement of 51818-51799 CTGCCGTCCCTTGAAGGCTA (SEQ ID NO:347)

Fragment 7

	Forward Primer 52932-52953 CCCTACCCTCAGGGATTTCTCA (SEQ ID NO:348)
5	Reverse Primer
	Complement of 53474-53455 CCCATTCTCCTCTCCGAGCA (SEQ ID NO:349)
	Fragment 8
10	Forward Primer 53280-53299 ATCAGCGTGGTGCGATGTGT (SEQ ID NO:350)
	Reverse Primer
	Complement of 53756-53737 ACCCAGCTCTCTGGGACACG (SEQ ID NO:351)
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	Fragment 9
	Forward Primer 53548-53568 GGGATGAGTTCCCAAGTGCAG (SEQ ID NO:352)
	Reverse Primer
20	Complement of 54048-54029 TGGCAAGCAGGCTTGAGAAG (SEQ ID NO:353)
	Fragment 10
	Forward Primer 53841-53860 CCCCAACCTGAGCCAGAAAC (SEQ ID NO:354)
25	Reverse Primer
	Complement of 54330-54311 TGTCCACAAGGGGGTCTGTG (SEQ ID NO:355)
	Fragment 11
20	Forward Primer 54083-54102 GGCTAGCAGTGGGGAAGAGG (SEQ ID NO:356)
30	Reverse Primer
	Complement of 54617-54600 ATTGCCAGGGGCAGGATG (SEQ ID NO:357)
	Fragment 12
35	Forward Primer
	54422-54441 CCCTGTCATGGCCAGTCCTT (SEQ ID NO:358) Reverse Primer
	Complement of 54929-54910 GCGACCCAGTGCCCTCTACT (SEQ ID NO:359)
40	Fragment 13
40	Forward Primer
	54694-54715 TGCATGTCCTCTTGTTGCTGAG (SEQ ID NO:360) Reverse Primer
	Complement of 55213-55194 CAATGACCACCCTCCCTGAA (SEQ ID NO:361)
45	From out 14
	Fragment 14 Forward Primer
	55273-55291 CGGCTGTCAAGGGGTGTTC (SEQ ID NO:362)
50	Reverse Primer Complement of 55769-55750 CCAAACCCAGACGCAAGTT (SEQ ID NO:363)

Analysis of Sequences for Polymorphic Sites

Sequences were analyzed for the presence of polymorphisms using the Polyphred program (Nickerson et al., Nucleic Acids Res. 14:2745-2751, 1997). The presence of a polymorphism was confirmed on both strands. The polymorphisms and their locations in the IL4Ra gene are listed in Table 3 below.

Example 1B

This example illustrates examination of the IL4R α gene for polymorphic sites in the following target regions: 1000 base pairs upstream of the ATG start codon; each of the exons, including approximately 100 base pairs on either side of the exon; and 500-1000 base paris downstream of the termination codon.

Amplification of the Target Regions

PCR primers, which were designed based on the nearly complete IL4R genomic sequence reported in the GenBank database (Accession No: AC004525), are set forth below:

Promoter 15

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Forward Primer

AAACCCCTGGACTCCAAGTGATCC (SEQ ID NO:364) 30094-30117

Reverse Primer

Complement of 30842-30819 AAGCGATTCTTCTGCCTCAGCCTC (SEQ ID NO:365)

PCR product 749 nt 20

Exon 1

Forward Primer

GGACAGTTGTTGTGTAGCTCACCC (SEQ ID NO:366) 30517-30540

25 Reverse Primer

Complement of 31409-31431 CTATGTTGCCCAAGCTGACCTC (SEQ ID NO:367)

PCR product 893 nt

Exon 2

Forward Primer 30

GCCCTCAACTTTGCCTGCAC (SEQ ID NO:368) 32871-32890

Reverse Primer

Complement of 33362-33340 AGTCCAGTATTCCAGCCGTATCC (SEQ ID NO:369)

PCR product 492 nt

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Exon 3

Forward Primer

TGATCGGGAAGCTGGAAGAGTC (SEQ ID NO:370) 35623-35644

Reverse Primer

Complement of 36070-36049 CGTTTCAAAGGTGGGAGGACTG (SEQ ID NO:371) 40

PCR product 448 nt

Exon 4

Forward Primer

CGACCAAAAATCTGGGTGGTG (SEQ ID NO:372) 45 37230-37250

Reverse Primer

Complement of 37668-37648 CAGGAAGCAAAGGGACTTGCC (SEQ ID NO:373)

PCR product 439 nt

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	Forward Primer 43307-43328 TCTTAAACATGGTGGGGTCAGC (SEQ ID NO:374)
5	Reverse Primer Complement of 43765-43745 CATGGAAATTGTGGGCTTGTG (SEQ ID NO:375) PCR product 459 nt
10	Exon 6 Forward Primer 46507-46528 ATGTGCAAGAGGGAGAGTGGTG (SEQ ID NO:376) Reverse Primer
	Complement of 46870-46849 TGACTGAGAGGACTGCAAAGGG (SEQ ID NO:377) PCR product 364 nt
15	Exon 7
	Forward Primer 49672-49695 GCCTGATCTCTGATGCCAAATAAG (SEQ ID NO:378) Reverse Primer
20	Complement of 49964-49983 TTTGCCATTCCAGAAGCCAG (SEQ ID NO:379) PCR product 293 nt
	Exon 8
	Forward Primer 51524-51546 GATCTGTGTGATGTCGAGGCTTG (SEQ ID NO:380)
25	Reverse Primer Complement of 51845-51825 GTCCACGTTTCCAGAACACCC (SEQ ID NO:381) PCR product 322 nt
30	Exon 9 Fragment 1 Forward Primer 52912-52933 CGAAATCCCAAAGACACAGACC (SEQ ID NO:382)
	S2912-52933 CGAAATCCCAAAGACACAGACC (SEQ ID NO:382) Reverse Primer Complement of 53722-53701 GAGTTGCTGAAGCTGCGGTAAG (SEQ ID NO:383) PCR product 811 nt
35	Exon 9 Fragment 2
	Forward Primer 53352-53374 GAAAAAGGGAGCTTCTGTGCATC (SEQ ID NO:384)
40	Reverse Primer Complement of 54172-54153 AACAAGGGGACAGGGACTGG (SEQ ID NO:385) PCR product 821 nt
	Exon 9 Fragment 3 Forward Primer
45	53854-53874 CAGAAACCTGGGAGCAGATCC (SEQ ID NO:386) Reverse Primer
	Complement of 54711-54688 GCAACAAGAGGACATGCACCTAAG (SEQ ID NO:387) PCR product 858 nt
50	Exon 9 Fragment 4 Forward Primer
	54266-54288 AAAGGTAGAGGACATGCCAAAGC (SEQ ID NO:388) Reverse Primer
55	Complement of 55007-54987 GGAGCAGCCAACAACTCGTTC (SEQ ID NO:389) PCR product 742 nt

These primer pairs were used in PCR reactions containing genomic DNA isolated from immortalized cell lines for a reference population of 70 human individuals. The PCR reactions were carried out under the following conditions:

5	Reaction volume	$= 20 \mu l$								
,	10 x Advantage 2 Polymerase reaction buffer (Clontech)	$= 2 \mu l$								
	100 ng of human genomic DNA	$= 1 \mu l$								
	10 mM dNTP	$= 0.4 \mu l$								
	Advantage 2 Polymerase enzyme mix (Clontech)	$= 0.2 \mu l$								
10	Forward Primer (10 µM)	$= 0.4 \mu l$								
10	Reverse Primer (10 μM)									
		=15.6µl								
	Water	•								
	Amplification profile:									
15	94°C - 2 min. 1 cycle									
	94°C - 30 sec.									

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70°C - 45 sec.

72°C - 1 min.

94°C - 30 sec. 64°C - 45 sec. 72°C - 1 min.

10 cycles

Sequencing of PCR Products

The PCR products were purified by Solid Phase Reversible Immobilization using the protocol developed by the Whitehead Genome Center. A detailed protocol can be found at http://www.genome.wi.mit.edu/sequencing/protocols/pure/SPRI_pcr.html.

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Briefly, five μ l of carboxyl coated magnetic beads (10 mg/ml) and 60 μ l of HYB BUFFER (2.5M NaCl/20% PEG 8000) were added to each PCR reaction mixture (20 μ l). The reaction mixture was mixed well and incubated at room temperature (RT) for 10 min. The microtitre plate was placed on a magnet for 2 min and the beads washed twice with 150 μ l of 70% EtOH. The beads were air dried for 2 min and the DNA was eluted in 25 μ l of distilled water and incubated at RT for 5 min. The beads were magnetically separated and the supernatant removed for testing and sequencing.

The purified PCR products were sequenced in both directions using the primer sets described previously or those listed, in the 5' to 3' direction, below.

Promoter Fragment 1

Forward Primer
30312-30334 GCTCATTTAATCCCCACAACACC (SEQ ID NO:390)
Reverse Primer

Complement of 30791-30769 CCACCACACCTGGCTAATTTTTG (SEQ ID NO:391)

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Promoter Fragment 2
Forward Primer
30529-30551 TGTAGCTCACCCTCTGGACTTTG (SEQ ID NO:392)
Reverse Primer
Complement of 30990-30971 AATATGCAACCCTCCCCTGC (SEQ ID NO:393)

Exon 1
Forward Primer
30824-30846 TGAGGCAGAAGAATCGCTTGAAC (SEQ ID NO:394)

10 Reverse Primer Complement of 31261-31240 ACTTGTCATTGGCTGTCCCCTC (SEQ ID NO:395)

Exon 2

Forward Primer

15 32880-32900 TTTGCCTGCACTGTGCTTTTG (SEQ ID NO:396)
Reverse Primer
Complement of 33234-33212 CCATACTCAGCATCCTGCACTCC (SEQ ID NO:397)

Exon 5

Forward Primer
43330-43349 AACGACAGCAACCAGGGTGG (SEQ ID NO:398)
Reverse Primer
Complement of 43704-43682 CAGCAGGTGTATCTAATGGCAGG (SEQ ID NO:399)

25 Exon 6 Forward Primer 46520-46541 AGAGTGGTGGGGAGATGAGGTG (SEQ ID NO:400)

Reverse Primer
Complement of 46857-46837 TGCAAAGGGGCAGACTAGAGG (SEQ ID NO:401)

30 Exon 7

Forward Primer 49708-49729 CGACCACTTTTATGGGAGGAGC (SEQ ID NO:402)

Reverse Primer

35 Complement of 49927-49905 CCAGGTGTTCTGAACCACACTTC (SEQ ID NO:403)

Exon 8
Forward Primer
51528-51550 TGTGTGATGTCGAGGCTTGTACC (SEQ ID NO:404)

40 Reverse Primer
Complement of 51779-51758 GAATGCAGGGAAGAGAGAGAGGCAG (SEQ ID NO:405)

Exon 9 Fragment 1 Forward Primer

45 53017-53038 GCCATCAGGACATGGTGATTTC (SEQ ID NO:406)
Reverse Primer
Complement of 53539-53518 TGAGCACTCGTACTTCCCGAAG (SEQ ID NO:407)

Exon 9 Fragment 2

50 Forward Primer
53378-53399 TGAGAGCAGCAGGGATGACTTC (SEQ ID NO:408)
Reverse Primer
Complement of 53948-53926 AAACTCCTGATAGCCACTGGTGG (SEQ ID NO:409)

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Exon 9 Fragment 3 Forward Primer AGATCCTCCGCCGAAATGTC (SEQ ID NO:410) 53869-53868 Reverse Primer TTACTCTTCTGAGATGCCCGAG (SEQ ID NO:411) Complement of 54583-54560

Exon 9 Fragment 4 Forward Primer TGGGCAGTGGCATTGTCTACTC (SEQ ID NO:412) 54334-54355

10 Reverse Primer Complement of 54769-54750 TTCCAGGAGGTGGCATTTCC (SEQ ID NO:413)

Sequencing reactions were performed using the Big-Dye terminator kit from PE Biosystems (Foster City, CA) according to the manufacturer's instructions. The sequencing products were analyzed on an ABI 477 automated sequencer (PE Biosystems, Foster City, CA).

Analysis of Sequences for Polymorphic Sites

Sequences were analyzed for the presence of polymorphisms using the Polyphred program (Nickerson et al., 14 Nucleic Acids Res. 2745-2751, 1997). The presence of a polymorphism was confirmed on both the strands. The polymorphisms and their locations in the IL4R α gene are listed in Table 3 below.

=	Nucleotide Position in	ic Sites Identified in the Nucleotide Position in	Reference	Variant	T
Polymorphic Site Number	GenBank Accession	Figure 1	Allele	Allele	Example
PS1	97137(Acc#AC004525)	32884	A	G	1B
PS2	97118(Acc#AC004525)	32903	С	T	1A
PS3	97060(Acc#AC004525)	32961	G	T	1A
PS4	96886(Acc#AC004525)	33135	G	С	1A
PS5 ^R	94272(Acc#AC004525)	35749	Α	G	1A,1B
	94258(Acc#AC004525)	35763	С	T	1A,1B
PS6 PS7	94251(Acc#AC004525)	35770	G	Α	1A
PS8	94204(Acc#AC004525)	35817	T	С	1A
PS9	94116(Acc#AC004525)	35905	С	T	1B
PS10	94077(Acc#AC004525)	35944	С	Т	1A
PS10 PS11	94063(Acc#AC004525)	35958	G	Α	1B
PS12	92691(Acc#AC004525)	37330	G	Α	1A
PS12 PS13	92548(Acc#AC004525)	37473	С	T	1A
PS13 PS14	92435(Acc#AC004525)	37586	C	T	1A
PS14 PS15	92430(Acc#AC004525)	37591	G	Α	1A
PS16	92417(Acc#AC004525)	37604	Α	T	1A
PS17	92377(Acc#AC004525)	37644	С	A	1A
PS18	92343(Acc#AC004525)	37678	С	T	1A
PS19	86575(Acc#AC004525)	43446	G	A	1A
PS20	86318(Acc#AC004525)	43703	Т	С	1A
PS21	77013(Acc#AC004525)		Α	С	1A
PS22	76922(Acc#AC004525)		С	Ť	1B
PS23	76868(Acc#AC004525)	<u> </u>	T	С	1A
PS24 ^R	76608(Acc#AC004525)		Α	С	1A
PS25	76565(Acc#AC004525)		G	T	1A, 1B
			Т	С	1A, 1B
PS26 ^R	76516(Acc#AC004525) 76514(Acc#AC004525)		Ċ	T	1A
PS27	76508(Acc#AC004525)	1	 	Ċ	1A
PS28			 	. C	1A, 1B
PS29 ^R	76300(Acc#AC004525)		C	 	1A
PS30	76106(Acc#AC004525)				1A, 1B
PS31 ^R	76080(Acc#AC004525)		A	G A	1A, 1B
PS32	76072(Acc#AC004525)		G	 	-
PS33	75784(Acc#AC004525)		C _	G	1A, 1B
PS34	75553(Acc#AC004525)		 	 	1A
PS35	75410(Acc#AC004525		T	 c	1A, 1B
PS36	75323(Acc#AC004525		T	 c	1A
PS37	75321(Acc#AC004525		C	 	1A
PS38	75280(Acc#AC004525		C	G	1A
PS39	75241(Acc#AC004525			G	1A
PS40	74938(Acc#AC004525		A G	A	1A
PS41	74879(Acc#AC004525			+	
PS42 ^R	74693(Acc#AC004525		G	A	1A
PS43 ^R	74591(Acc#AC004525		С	T	1A
PS44	74482(Acc#AC004525	55539	С	T	1A
PS45	74263(Acc#AC004525) 55758	G	A	1A

^RPreviously reported in the literature

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Example 2

This example illustrates analysis of the IL4R α polymorphisms identified in the Index Repositories for human genotypes and haplotypes for all polymorphic sites except PS1, PS9, PS11, PS21, PS22, and PS23.

A sampling of different genotypes containing these polymorphisms that were observed in these reference populations are shown in Table 4 below, with the haplotype pair indicating the combination of haplotypes determined for the individual using the haplotype derivation protocol described below. In Table 4, homozygous positions are indicated by one nucleotide and heterozygous positions are indicated by two nucleotides. Missing nucleotides in any given genotype in Table 4 can typically be inferred based on linkage disequilibrium and/or Mendelian inheritance.

44 ပပြ ග ⋖ ပြ ഗ < 0 0 0 ര ပ ഠ O တ PS G മ G/A ග G G PS 37 PS 36 PS 35 PS 34 ര ග G G ဖြ ഗ PS 32 ľО G ග ര PS 31 O PS 30 PS 29 PS 28 PS 27 PS 26 G F 5 ഗ O PS 25 Table 4. Genotypes and HAP Pairs Observed for the IL4R α Gene PS 24 PS 20 တ ര G Ö G ග PS 19 တပြာ စ O. PS 18 PS 17 PS 16 တတ ഗ G PS 15 G ပ ပ ပ O G ගර G ပြုပ ဖြ ტ G G G ပြ ග ပပ ଠାଠ G ഠ PS 6 8€ G/A ග G G/A ഗ G G ഗ G G ပြာ ပ G G G G PS G ဗြ G G G ල G 9 ပ ပြ G ග ග ග ග ഗ G တ ტ ග ပ ပ ပပြ ပ PS C

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The haplotype pairs shown in Table 4 were estimated from the unphased genotypes using an extension of Clark's algorithm (Clark, A.G. (1990) *Mol Bio Evol* 7, 111-122), as described in U.S. Provisional Patent Application filed April 19, 2000 and entitled "A Method and System for Determining Haplotypes from a Collection of Polymorphisms". In this method, haplotypes are assigned directly from individuals who are homozygous at all sites or heterozygous at no more than one of the variable sites. This list of haplotypes is then used to deconvolute the unphased genotypes in the remaining (multiply heterozygous) individuals.

By following this protocol, it was determined that the Index Repositories examined herein and, by extension, the general population contains the 53 human IL4R α haplotypes shown in Table 5 below.

Table 5A. Haplotypes Observed for the IL4Rlpha Gene

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PS	43	၁	ပ	ပ	ပ	ပ	ပ	의	-	ပ		익	ပ	닏	늬	ပ	ပ	의	의	늬			-	-	0	-		\dashv			\dashv	ပ	-	_	의
PS	42	A	ပ	⋖	ပ	⋖	۷	ପ	⋖ :	⋖ .	< -	<	9	⊴	≤	ග	ଠ	ଠ		≤	\dashv		-+	-	۷	⋖	4	Ϥ	≤	≤	≤	≤	۷		의
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PS	39	၁	၁	ပ	ပ	ပ	ပ	ပ	ပ	ပြ	ن ا	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	이	ပ	의	ပ	ပ	ଠ	이	ပ	의	이	0	의
PS	38	၁	ပ	C	ပ	ပ	၁	ပ	ပ	ပ	-	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	0	의	ပ	ပ	ပ	이	ပ	ပ	의	이	<u>ပ</u> ု	의
PS	37	T	⊢	_	⊢	⊢	⊢	⊢	H	-	\vdash	ပ	-	ပ	\vdash	\vdash		⊢	Ы	 		⊢	⊢		니	ပ	⊢	⊢	ပ	ပ		<u>- </u>	-	븨	늬
PS		-	F	ļ.	ပ	⊢	⊢	F		-		ပ	-	-	ပ	\vdash	-	⊢	⊢	ပ		⊢	$\vdash \mid$		니	⊢	0	⊢	⊢	<u>-</u>	⊢	니	Ы		
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PS	3	ပ	ပ	ပ	ပ	၁	၁	၁	ပ	0	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	С	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	이	ပ	의	ပ	ပ
Sd	7	O	O	ပ	ပ	G	9	9	٧	0	ပ	O	ပ	ပ	ပ	ß	ග	ပ	ပ	ပ	ග	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	G	۷	0	0
Sd	-	⋖	A	⋖	Ö	Ö	Α	ပ	ပ	O	ပ	ပ	ပ	O	ပ	٧	۷	9	Α	၁	ပ	9	٧	∢	٧	ග	O	Α	O	ပ	ଠ	ပ	ပ	ပ	⋖
Sd	0	ပ	ပ	ပ	ပ	ပ	ပ	ပ	၁	ပ	ပ	ပ	ပ	ပ	ပ	ပ	C	С	С	С	ပ	၁	0	O	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	늬
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S.d	7	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ن	ပ	ပ	ပ	ပ	ပ	၁	C	ပ	ပ	Ţ	L	ပ	ပ	С	ပ	ပ	၁	၁	S	၁	ပ	ပ	ပ	ပ	ပ
8	9	F	F	-	-	ပ	⊢	ပ	F	ပ	\vdash	-	⊢	-	┙	T	⊢	ပ	⊢	} —	T	-	⊢	_	-	_	1	Ţ	T	⊥	Η	_	\vdash	၁	느
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Table 5B. Haplotypes Observed for the IL4Rlpha Gene

PS	45	٧	Ö	Α	ව	Ŋ	Α	Α	Α	ග	A	ව	۷	ပ	Α	Α	Y.	9	9	Α
PS	44	၁	C	ပ	С	ပ	၁	С	С	ပ	C	၁	၁	ပ	C	C	0	С	Э	C
PS	43	၁	C	Ţ	ပ	ပ	၁	С	C	ပ	Ċ	С	C	၁	С	C	Ţ	၁	Э	⊢
PS	42	Α	G	A	G	ပ	٧	Α	Α	ပ	Α	G	٧	٧	Α	Α	٧	G	Α	А
PS	41	ß	G	G	G	9	ß	G	ပ	ග	G	G	G	ß	G	G	9	9	9	9
PS	40	G	Α	G	A	A	G	O	ပ	٧	G	Α	G	G	G	G	9	٧	9	9
PS	39	С	C	၁	၁	ပ	ပ	ပ	ပ	ပ	C	၁	၁	၁	С	C	Э.	၁	Э	ပ
PS	38	၁	၁	၁	С	ပ	C	ပ	ပ	ပ	Ö	Э	၁	၁	၁	С	С	ပ	၁	၁
PS	37	Ţ	Ė	T	┙	⊢	1	С	T	⊢	Ţ	T	1	⊢	⊢	_	Ţ	Н	⊢	⊢
PS	36	T	ပ	Τ	Ţ	!	Τ	С	H	-	T	-	⊢	⊢	⊢	-	၁	⊢	⊢	ပ
PS	35	Τ	Τ	Ţ	Ţ	⊢	⊢	⊢	Η:	-	Ξ	Η.	⊢	1	┙	_	T	⊢	\vdash	⊢
PS	34	Ţ	Τ	G	Ţ	⊢	⊢	ව	⊢		_	Τ.	_	⊢	⊢	9	9	⊢	⊢	ග
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PS	32	9	9	А	9	ව	9	G	ပ	ပ	G	9	9	9	၅	G	9	၅	9	ග
PS	31	٧	ß	Э	9	Α	Α	ව	٨	ග	A	٧	Ä	Э	Α	၅	G	٧	G	9
PS	30	Э	Ö	<u>)</u>	Э	၁	ပ	ပ	C	ပ	ပ	၁	၁	၁	၁	Ċ	ပ	၁	၁	၁
PS	29	L	Ţ	1	_	⊢	⊥	⊢	⊢	၁	⊥	Ι.	⊢	ပ	1	\perp	⊢	_	၁	—
PS	28	1	1	၁	T	T	T	⊢	T	၁	1	⊢	H	၁	⊥	၁	ပ	1	၁	ပ
PS	27	Э	Э	С	၁	၁	၁	Ö	၁	S	၁	၁	0	ပ	၁	၁	ပ	C	၁	၁
PS	26	⊥	·l	⊢	⊢	⊢	⊥	⊢	⊥	၁	Ţ	۰	⊢	ပ	⊢	-	⊥	⊢	၁	┪
PS	25	ອ	9	-	၅	9	9	9	ව	l	ပ	9	၅	⊢	9	⊢	⊢	ර	۲	-
Sd	24	٧	٧	ပ	⋖	Α:	٧	٧	٧	၁	⋖	۷	A	Ü	٧	C	၁	٧	၁	၁
Sd	20	⊢	ပ	ပ	ပ	ပ	၁	၁	⊥	၁	ပ	⊢	⊢	ပ	ပ	၁	ပ	⊥	၁	၁
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Sd	8	ပ	<u> -</u>	-	ပ	⊢	ပ	0	ပ	ပ	<u></u>	ပ	ပ	-	-	၁	ပ	ပ	-	ပ
PS	17	ပ	O	ပ	ပ	ပ	ပ	၁	Ö	ပ	ပ	ပ	ပ	O	ပ	C	ပ	0	ပ	ပ
PS	16	<	۷	4	⋖	۷	⋖	٧	⋖	٧	⋖	⋖.	⋖	<	٧	٧	٧	٧	∢.	۲.
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PS	13	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	0	ပ	0	ပ	⊢	ပ	ပ	ပ	ပ
PS	12	၅	Ø	ပ	O	O.	0	O	O	ပ	9	ပ	9	O	ග	Ö	O	Ö	S	
PS	19	ပ	ပ	0	ပ	ပ	ပ	ပ	ပ	ပ	-	 -	⊨	⊢	<u> </u>	⊢	-	ပ	上	၁
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PS	7	၅	ပ	ပ	Ö	O	ග	Ö	٧	⋖	ပ	ပ	ပ	9	O	O	ß	ပ	ပ	ပ
PS	9	ပ	ပ	ပ	၁	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	ပ	၁	ပ	၁	ပ	ပ	⊢
PS	2	⋖	⋖	٧	٧	⋖	⋖	Ö	⋖	⋖	9	၅	Ö	ග	g	ပ	G	٧	ပ	G
PS	4	ပ	ග	G	O	g	(5)	ß	O	ပ	ပ	9	ဗ	9	Ö	0	G	9	9	G
PS	ო	ပ	O	9	O	9	ဗ	ပ	O	9	၅	ග	ß	9	O	၅	O	9	<u> -</u>	9
PS	7	ပ	ပ	ပ	O	ပ	ပ	⊢	ပ	ပ	0	ပ	ပ	0	F	၁	ပ	ပ	L	۲
₽	≘	35	36	37	38	39	40	41	42	43	44	45	46	47	48	49	20	51	52	53

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In view of the above, it will be seen that the several advantages of the invention are achieved and other advantageous results attained.

As various changes could be made in the above methods and compositions without departing from the scope of the invention, it is intended that all matter contained in the above description and shown in the accompanying drawings shall be interpreted as illustrative and not in a limiting sense.

All references cited in this specification, including patents and patent applications, are hereby incorporated in their entirety by reference. The discussion of references herein is intended merely to summarize the assertions made by their authors and no admission is made that any reference constitutes prior art. Applicants reserve the right to challenge the accuracy and pertinency of the cited references.